

PATENT COOPERATION TREATY

PCT

NOTIFICATION OF ELECTION

(PCT Rule 61.2)

From the INTERNATIONAL BUREAU

To:

Commissioner
US Department of Commerce
United States Patent and Trademark
Office, PCT
2011 South Clark Place Room
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Arlington, VA 22202
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in its capacity as elected Office

Date of mailing (day/month/year)

02 May 2001 (02.05.01)

International application No.

PCT/EP00/08371

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5265/00/WO-Ab

International filing date (day/month/year)

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Priority date (day/month/year)

30 August 1999 (30.08.99)

Applicant

SEELA, Frank et al

1. The designated Office is hereby notified of its election made:



in the demand filed with the International Preliminary Examining Authority on:

03 February 2001 (03.02.01)



in a notice effecting later election filed with the International Bureau on:

2. The election



was



was not

made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO
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99116767.7 30 August 1999 (30.08.1999) EP (84) Designated States (regional): European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).
- (71) Applicant (for all designated States except US): ROCHE DIAGNOSTICS GMBH [DE/DE]; D-68298 Mannheim (DE).
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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



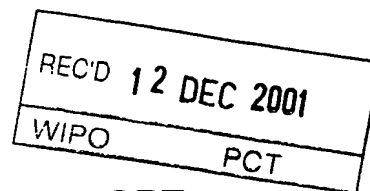
WO 01/16149 A2

(54) Title: 2-AZAPURINE COMPOUNDS AND THEIR USE

(57) Abstract: Within oligonucleotides 2-azapurine and especially 2-azaadenine bases form specifically base pairs with guanine. This base pair is of analogous stability as an adenine-thymine but less stable than a guanine-cytosine base pair. Therefore, the incorporation of 2-azaadenine residues into oligonucleotides instead of cytosine leads specifically to hybridization complexes with nucleic acids with homogenous stability. This is useful for the adaptation of the stabilities of different oligonucleotide sequences in all kinds of hybridization techniques, for example in oligomer chip technology.

PATENT COOPERATION TREATY

PCT



INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)


Applicant's or agent's file reference 5265/00/WO-Ab	FOR FURTHER ACTION See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)	
International application No. PCT/EP00/08371	International filing date (day/month/year) 28/08/2000	Priority date (day/month/year) 30/08/1999
International Patent Classification (IPC) or national classification and IPC C07H19/00		
Applicant ROCHE DIAGNOSTICS GMBH		

1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.
2. This REPORT consists of a total of 8 sheets, including this cover sheet.
 - ☐ This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).

These annexes consist of a total of sheets.

3. This report contains indications relating to the following items:

- I ☒ Basis of the report
- II ☐ Priority
- III ☐ Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
- IV ☐ Lack of unity of invention
- V ☒ Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
- VI ☒ Certain documents cited
- VII ☐ Certain defects in the international application
- VIII ☒ Certain observations on the international application

Date of submission of the demand 03/02/2001	Date of completion of this report 07.12.2001
Name and mailing address of the international preliminary examining authority:  European Patent Office - P.B. 5818 Patentlaan 2 NL-2280 HV Rijswijk - Pays Bas Tel. +31 70 340 - 2040 Tx: 31 651 epo nl Fax: +31 70 340 - 3016	Authorized officer Scott, J Telephone No. +31 70 340 2206



**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT**

International application No. PCT/EP00/08371

I. Basis of the report

1. With regard to the **elements** of the international application (*Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to this report since they do not contain amendments (Rules 70.16 and 70.17)*):

Description, pages:

1-48 as originally filed

Claims, No.:

1-35 as originally filed

Drawings, sheets:

1/6-6/6 as originally filed

2. With regard to the **language**, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

- ☐ the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
- ☐ the language of publication of the international application (under Rule 48.3(b)).
- ☐ the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

- ☐ contained in the international application in written form.
- ☐ filed together with the international application in computer readable form.
- ☐ furnished subsequently to this Authority in written form.
- ☐ furnished subsequently to this Authority in computer readable form.
- ☐ The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
- ☐ The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:

- ☐ the description, pages:
- ☐ the claims, Nos.:

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT**

International application No. PCT/EP00/08371

☐ the drawings, sheets:

5. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):
(Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)

6. Additional observations, if necessary:

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N)	Yes:	Claims	5,6,8,9,15-18,26-35
	No:	Claims	1-4,7,10-14,19-25
Inventive step (IS)	Yes:	Claims	
	No:	Claims	1-35
Industrial applicability (IA)	Yes:	Claims	1-35
	No:	Claims	

2. Citations and explanations
see separate sheet

VI. Certain documents cited

1. Certain published documents (Rule 70.10)

and / or

2. Non-written disclosures (Rule 70.9)

see separate sheet

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:
see separate sheet

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT - SEPARATE SHEET**

International application No. PCT/EP00/08371

Re Item V

Reasoned statement under Rule 66.2(a)(ii) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

Cited documents :

Reference is made to the following documents :

D1 : M.ACEDO ET AL.: "Synthesis and Biophysical and Biological Properties of Oligonucleotides Containing 2-Aza-2'-deoxyinosine" JOURNAL OF ORGANIC CHEMISTRY, vol. 60, no. 20, 1995, pages 6262-6269, XP000971913 60

D2 : R.ERITJA ET AL.: "Preparation of Oligonucleotides Containing Non-Natural Base Analogues." NUCLEOSIDES AND NUCLEOTIDES, vol. 14, no. 3-5, 1995, pages 821-824, XP000971912

D3 : D.FERNANDEZ-FORNER ET AL.: "Preparation of Oligonucleotides Containing dAICA Using an Unexpected Side-Reaction Observed on a Projected Derivative of 2-Aza-2'-Deoxyinosine." TETRAHEDRON, vol. 47, no. 42, 1991, pages 8917-8930, XP000978346

D4 : L.L.BENNETT ET AL.: "Nucleosides of 2-Aza-Purines - Cytotoxicities and Activities as Substrates for Enzymes Metabolizing Purine Nucleosides." BIOCHEMICAL PHARMACOLOGY, vol. 25, no. 5, 1976, pages 517-521, XP000978454

D5 : MONTGOMERY J A ET AL: "NUCLEOSIDES OF 2-AZAPURINES. 7H-IMIDAZO(4,5-D-1,2,3-TRIAZINES" JOURNAL OF MEDICINAL CHEMICAL SOCIETY. WASHINGTON,US, vol. 18, no. 6, 1975, pages 564-567, XP000971950 ISSN: 0022-2623

D6 : J.C.BUSSOLARI ET AL.: "The Synthesis and Biological Evaluation of 4-p-Nitrobenzylthio-v-triazolo(4,5-d)pyridazine and Imidazo(4,5-d)pyridazine Ribosides as Potential Nucleoside Transport Inhibitors." BIOORGANIC &

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT - SEPARATE SHEET**

International application No. PCT/EP00/08371

MEDICINAL CHEMISTRY, vol. 4, no. 10, 1996, pages 1725-1731, XP000978366

D7 : J.A.CARBON: "Synthesis of Some Imidazo(4,5-d)pyridazines and Imidazo(4,5-d)triazolo(4,3-b)pyridazines." JOURNAL OF ORGANIC CHEMISTRY., vol. 25, no. 4, 3 May 1960 (1960-05-03), pages 579-582, XP002160096 AMERICAN CHEMICAL SOCIETY. EASTON., US ISSN: 0022-3263

D8 : Z.KAZIMIERCZUK ET AL.: "Stereoselective Synthesis of 2-Azapurine 2'-Deoxy-B-D-ribonucleosides by Nucleobase-Anion Glycosylation." LIEBIGS ANNALEN DER CHEMIE., no. 7, 1990, pages 647-651, XP002160097 VERLAG CHEMIE GMBH. WEINHEIM., DE ISSN: 0170-2041

Novelty

Document D1 discloses p.6263, figure 1, oligonucleotides containing 2-Aza-2'-deoxyinosine - these compounds can circumvent the problem of degeneracy of the genetic code by being able to bind to any of the four naturally occurring nucleobases. Compounds 8,9,14,15,17,10,16,11,12 (fig.3, p.6264) ALL fall within the scope of claim 13. Moreover, compound 13 (fig.3, p.6264) falls within the scope of claim 14.

D2 details the preparation of oligonucleotides containing non-natural base analogues. More specifically it contains 2-azahypoxanthine and 2-aza-2'-deoxyinosine. This is done in an attempt to improve the base-pairing properties of hypoxanthine.

D3 teaches the introduction of 2-aza-2'-deoxyinosine into oligonucleotides.

Finally, D4-D8 all describe compounds falling within the scope of claim 13.

Thus the subject-matter of claims 1-4,7,9-14,16,19-25 lack novelty in the sense of Article 33(2) PCT.

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT - SEPARATE SHEET**

International application No. PCT/EP00/08371

Inventive Step

In so far as the claims are novel, the claimed compounds only differ from those of the closest prior art in the selection of a list of interchangeable variables.

The problem to be solved by the present application is regarded as the provision of further nucleic acid binding compounds with a base structure of formula (I).

The person skilled in the art, when wishing to solve this problem is provided with many incentives from the prior art to utilise different variables (see docs D1-D8) with the expectation that the resultant compounds would still bind to nucleic acids.

Thus the subject-matter of claims 1-35 cannot be regarded as involving an inventive step in the sense of Article 33(3),PCT.

support

There is not much support for the scope of claims on file. Indeed, the examples cover only a tiny fraction of the claimed scope. Furthermore, many of the terms used to define the scope are not clear (see section VIII). Finally, it is doubted whether all, or even many, of the above compounds can solve the claimed problem (even drafted in a broad way - see above).

All the examples (which fall within the scope of the product claims) refer to similar types of compounds - namely :

2'-deoxy-B-D-erythropentofuranosyl with imidazo-triazine bases - this certainly does not give support to the compounds across the whole range of those claimed.

note : examples dealing with compounds which are non-claimed intermediates or starting materials should be designated as "preparative examples".

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT - SEPARATE SHEET**

International application No. PCT/EP00/08371

Re Item VI

Certain documents cited

T.SUGIYAMA ET AL.: "2-Aza-2'-Deoxyadenosine : Synthesis, Base-Pairing Selectivity, and Stacking Properties of Oligonucleotides." CHEMISTRY A EUROPEAN JOURNAL, vol. 6, no. 2, 17 January 2000 (2000-01-17), pages 369-378, XP000971911

CHEMICAL ABSTRACTS, vol. 133, no. 7, 14 August 2000 (2000-08-14) Columbus, Ohio, US; abstract no. 89731z, F.SEELA ET AL.: "Synthesis, Base Pairing and Stacking Properties of Oligonucleotides Containing 2-Aza-2'-deoxyadenosine." page 700; column 1; XP002160098 & COLLECT. SYMP. SER., no.2, 1999,124-128

Re Item VIII

Certain observations on the international application

i). With respect to the claims as a whole, there are too many independent claims - at least fourteen (1,13,14,19,20,22,23,24,26,28,29,30,34,35) thus causing the application to lack clarity and conciseness.

ii). Claim 1 is unclear for the reasons outlined below :

- the definition "A nucleic acid binding compound" attempts to define the scope of protection sought in terms of a result to be achieved - as does "capable of base pairing"
- the term "protecting group" is not well defined, since it is not clear what sort of structure requires protection
- the term "reporter group" (p.50, l.15) is vague and seeks to extend the scope of protection sought in an imprecise manner
- alkynyl appears to be an error (p.50, l.14)

**INTERNATIONAL PRELIMINARY
EXAMINATION REPORT - SEPARATE SHEET**

International application No. PCT/EP00/08371

most of these problems recur at various places in the claims.

iii). Furthermore, the following clarity objection is noted with respect to claim 11 :

- - the term "solid phase" (p.55, l.12) is vague and seeks to extend the scope of protection sought in an imprecise manner

iv). There are numerous problems throughout the claims with respect to the consistency of the use of variables - for example - claim 15 is dependent on claim 14, which details OR13, whereas claim 15 calls it OR31; claim 18 then refers to claim 5, which should be claim15, and R31 (R13?).

(19) World Intellectual Property Organization
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[DE/DE]; In der Klappe 9, 34399 Oberweser (DE). **HEINDL, Dieter** [DE/DE]; Waldschmidtstrasse 1, 82327 Tutzing (DE). **BERGMANN, Frank** [DE/DE]; Faltergatter 5, 82393 Iffeldorf (DE).

(74) Common Representative: **ROCHE DIAGNOSTICS GMBH**; Patent Dept., D-82372 Penzberg (DE).

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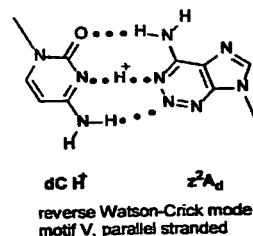
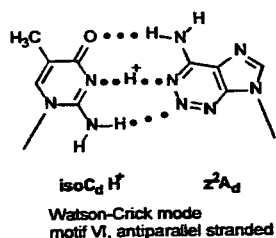
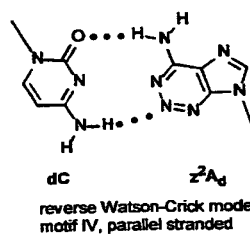
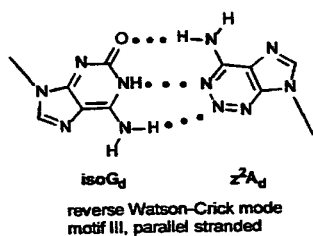
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2 August 2001

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(54) Title: **2-AZAPURINE COMPOUNDS AND THEIR USE**



(57) Abstract: Within oligonucleotides 2-azapurine and especially 2-azaadenine bases form specifically base pairs with guanine. This base pair is of analogous stability as an adenine-thymine but less stable than a guanine-cytosine base pair. Therefore, the incorporation of 2-azaadenine residues into oligonucleotides instead of cytosine leads specifically to hybridization complexes with nucleic acids with homogenous stability. This is useful for the adaptation of the stabilities of different oligonucleotide sequences in all kinds of hybridization techniques, for example in oligomer chip technology.

WO 01/16149 A3

5 2-Azapurine compounds and their use

The present invention is directed to a nucleic acid binding compound comprising 2-azapurines, a compound useful for the preparation of such compound, a binding
10 product of this nucleic acid binding compound with a nucleic acid, a method for the determination of a nucleic acid using said compound, and several uses of 2-azapurine compounds.

Some 2-azapurine compounds are known in the art. In Chemistry of Nucleosides
15 and Nucleotides (Vol. 2, p. 288-297, and p. 319) there are shown some examples of 2-azapurine nucleosides. However, there is no disclosure on 7-deaza-2-azapurine nucleosides.

In Biochimica et Biophysica Acta, 520 (1978), p. 441-451, there is disclosed the
20 enzymatic synthesis of 2-azaadenosine-5'-diphosphate and 2-aza inosine-5'-diphosphate and their polymerization to homopolymers using E.coli polynucleotide phosphorylase. This method is complex and it is not capable of producing mixed sequences. Furthermore, there was reported on the capability of the homopolymers to form double- and triple-stranded complexes with other homopolymers. Those
25 compounds only containing 2-azaadenosine as base have proved to be UV sensitive and labile.

In J. Org. Chem. 1995, 60, 6262-6269, there is disclosed the synthesis and biophysical and biological properties of oligonucleotides containing 2-aza-2'-deoxyinosine.
30 There is disclosure on a 2-aza inosine nucleoside, which is modified by a particular photochemically cleavable protecting group at one of the ring nitrogen atoms. However, as can be seen from table 1 on page 6266, I^{Az} does not differentiate well between the natural nucleobases. As can be seen further, the stability of the base pair I^{Az}/G is 13 ° less than the regular base pair C/G and 9 ° less than the base pair A/T. The replacement of C
35 by I^{Az} would therefore not be suitable for mimicking the stability of a A/T base pair.

In Liebigs Ann. Chem. 1990, 647-651, there are disclosed 2-azaadenine and a nucleoside of methylthioimidazotriazine as well as the corresponding methoxy compound. No phosphates or phosphoramidites are disclosed. Furthermore, there is no disclosure on how to prepare oligonucleotides containing 2-aza-adenosine at specific positions. Their hybridization behaviour is not disclosed. Furthermore, there is no disclosure on 2-aza-2'-deoxyadenosinetriphosphate.

The present invention is particularly useful in nucleic acid determinations, for example in analytics, especially in the field of health care. Nucleic acids have been found to be useful analytes for the determination of the presence or absence of genes or micro-organisms in human body fluids, food or environment. Nucleic acid analysis has found widespread use after the introduction of nucleic acid amplification, like the Polymerase Chain Reaction (PCR, see US-A-4,683,202). Thus, sufficient amounts of nucleic acids are available from each sample. The nucleic acids can be determined from this pretreated sample using a variety of different techniques, dependent from the particular purpose. Most assays require the use of a probe which is either immobilized or immobilizable or is labelled by attachment of one or more reporter groups. A reporter group has the characteristics to be itself capable to be determined or it can be reacted with reagents that make the probe determinable via said reporter group. Thus, for example, probes that are labelled by reporter groups can be determined, as can be hybrids that contain the probe and a nucleic acid to be determined. In case of immobilized probes, the hybrid between the probe and the nucleic acid to be determined is determined at the solid phase to which the probe is bound. In a particular form of assays, not only one nucleic acid having a specific sequence, but a large number of nucleic acids of different sequence is determined. For this purpose, the probes are immobilized in tiny spots in an array on a flat surface such as a glass chip (EP-A-0 476 014 and TIBTECH (1997), Vol. 15, 465-469).

The basic principle of using oligonucleotide arrays was first proposed in the late 1980s when the concept of determining a DNA sequence by hybridization to a comprehensive set of oligonucleotides (SBH, sequencing by hybridization) was developed.

There are many proposals to include modified or non-natural heterocyclic groups instead of the natural nucleobases. Examples of such non-natural groups are 7-deaza-dGTP which, when introduced into a nucleic acid replacing dGTP reduces band

compressing in sequencing gels (EP-B-0 286 028).

Nucleic acid determinations generally suffer from the problem that the base pairing possibilities between the natural bases A and T and C and G have different stability.

5 This can be attributed to the different capability of the bases to form hydrogen bonding. Thus, the dA-dT- base pair has two hydrogen bridges, while the dG-dC- base pair has 3 hydrogen bridges. This results in different melting temperatures (T_m) of hybrids, depending on the GC content. The higher the GC content, the higher the T_m . In routine nucleic acid analysis, however, there would be the wish to equalize the T_m for
10 nucleic acids of the same length, or even independent from the length of the nucleic acid or the binding region in order to be in the position to apply similar hybridization conditions for all assays. This is particularly necessary for assays using arrays, as on such arrays the hybridizing conditions for each probe must be identical.

15 One solution was the use of low hybridization temperatures. Under such conditions, many nucleic acids having a low degree of base sequence complementarity will bind to the probe. This is called unspecific binding which does not allow discrimination between similar sequences.

20 Another proposal was directed to the use of chemical reagents in the hybridization mixture, for example the addition of tetramethylammonium chloride (TMAC). This reagent reduces the difference between the stability of dG-dC and dA-dT basepairs but the effect is insufficient for short oligonucleotides. Further the addition of salts such as TMAC may not be wellcome as it complicates optimization of the assay.

25 Another proposal was directed to the use of different concentrations of each different (immobilized) probe in one assay. This was found to be technically complex if not impossible on a chip surface.

30 As a further option the substitution of ribonucleotides in an oligonucleotide composed of deoxyribonucleotides, and vice versa was applied for the adaptation of DNA stability, Hoheisel (1996), Nucleic Acids Res. 24, 430-432.

All proposals known now have some disadvantages. Therefore, there is still a need to
35 provide probes the T_m of which is not very dependent from their GC content.

FIG 1 shows how z^2A_d is capable of base pairing with a dG in another strand of a nucleic acid.

- 5 In FIG 2 there are shown different compounds of the invention, like 2-aza-2'-deoxyadenosine (2), the corresponding triphosphate (11), a phosphoramidite for the introduction of 2-aza-2'-deoxyadenosine into oligonucleotides during conventional chemical automated synthesis (10 b) and a H-phosphonate (12).
- 10 FIG 3 shows compounds useful in the present invention. The purine numbering for compound 2 and the systematic numbering for compound 3 is given.

FIG 4 shows a route for synthesis of 2-aza-2'-deoxy-adenosine.

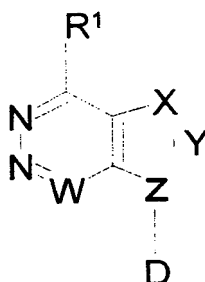
- 15 FIG 5 shows compounds of the invention.

FIG 6 shows a comparison of the binding of z^2A_d with dG and dG with isoG_d in antiparallel binding.

- 20 FIG 7 shows that z^2A_d can also bind in a parallel mode to isoG_d, can bind to dC in parallel mode when protonated, and to isoC_d in antiparallel mode if protonated.

- The subject of the present invention is a nucleic acid binding compound comprising a backbone, said backbone having attached heterocyclic groups capable of base pairing to
- 25 natural nucleobases at least one of said heterocyclic groups being one of the naturally occurring nucleobases characterized in that at least one other of said heterocyclic groups is a group of the general formula I

Formula I



wherein

5 W is selected independently from X, Y and Z from the group consisting of N and CR²,

 Z is selected from the group consisting of N and C with the proviso that

- if Z is N, then

10

 X independently from W and Y is selected from the group consisting of N and CR³, and

 Y independently from W and X is selected from the group consisting of N and CR⁴,

15

 and the bond between X and Y is a double bond and the bond between Y and Z is a single bond, and

- if Z is C, then

20

 X is NR³³, and

 Y is selected from the group consisting of N and CR⁴ and the bond between Z and Y is a double bond and the bond between X and Y is a single bond,

25

 R¹, R², R³ and R⁴ are independently selected from the group consisting of -H, -halogen, -OR¹³, -SR¹⁹, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₂-C₁₀)-alkynyl, -NO₂, -NR⁵R⁶, -cyano, and -C(=O)R¹¹,

R^{11} is selected from the group consisting of -OH, $-(C_1-C_6)$ -alkoxy, $-(C_6-C_{22})$ -aryloxy, and NHR^{12} ,

R^5 , R^6 , R^{12} , R^{13} , R^{19} and R^{33} are selected independently from the group consisting of -H, $-(C_1-C_{10})$ -alkyl, $-(C_2-C_{10})$ -alkenyl, $-(C_2-C_{10})$ -alkynyl, $-(C_6-C_{22})$ -aryl, a protecting group and a reporter group,

r and s are independently of each other an integer of 1 to 18,

D is the position of attachment of the group to the rest of the nucleic acid binding compound, and

said alkyl, alkenyl and alkynyl being unsubstituted or substituted by one or more moieties selected from the group consisting of -halogen, -SH, $-S-(C_1-C_6)$ -alkyl, $-(C_1-C_6)$ -alkoxy, -OH, $-NR^5R^6$, $-COR^{11}$, $-NH-CONR^5R^6$, $-NH-CSNR^5R^6$ and $-[O-(CH_2)_r]_s-NR^5R^6$.

By way of example, in the following there is given an explanation of the advantageous property of the compounds of the invention by showing them at the example of 2-aza-2'-deoxyadenosine ($2, z^2A_d$) which forms specifically stable base pairs with 2'-deoxyguanosine (dG) but much less stable base pairs with 2'-deoxythymidine (dT), 2'-deoxycytidine (dC) and 2'-deoxyadenosine (dA). The new base pair (z^2A_d -dG) is of analogous stability as a regular dA-dT base pair.

In order to equalize T_m , in a nucleic acid binding compound one or more C in a strand complementary to a G in the nucleic acid to be determined could be replaced by a z^2A_d . The oligonucleotide would then bind specifically to the target sequence containing dG opposite to z^2A_d but with the stability of a dA-dT and not a dG-dC base pair. This general principle of course is not limited to z^2A_d , as bases showing the same characteristics in the 6-membered ring would be expected to have the same properties based on the above explanation due to their containing the 2-azapurine structure. Particularly, the farther the part of the heterocyclic group from the part participating in the base pairing, the more tolerant will the oligomer be over modifications in the chemical structure, for example the attachment of groups to this part of the heterocyclic rings. In the following, when reference is made to the modified base (of the invention), there is made reference to a heterocyclic group according to general formula I.

The broken line in formula I indicates that there are several possibilities, depending upon the definitions of X, Y and Z, to localize a double bond. It is apparent to the man skilled in art that the choice of a specific definition of Z will require a double bond to be either between Z and Y or between X and Y. It is further evident that there will be not
5 each a double bond between X and Y and Y and Z.

Halogen means a fluoro, chloro, bromo or iodo group. The most preferred halogen groups are -Cl and -Br.

10 Alkyl groups are preferably chosen from alkyl groups containing from 1 to 10 carbon atoms, either arranged in linear, branched or cyclic form. The actual length of the alkyl group will depend on the steric situation at the specific position where the alkyl group is located. If there are steric constraints, the alkyl group will generally be smaller, the methyl and ethyl group being most preferred. All alkyl, alkenyl and alkynyl groups can
15 be either unsubstituted or substituted. Substitution by hetero atoms as outlined above, will help to increase solubility in aqueous solutions.

Alkenyl groups are preferably selected from alkenyl groups containing from 2 to 10 carbon atoms. For the selections similar considerations apply as for alkyl groups. They
20 also can be linear, branched and cyclic. The most preferred alkenyl group is the ethylene group.

Alkynyl groups have preferably from 2 to 10 carbon atoms. Again, those carbon atoms can be arranged in linear, branched and cyclic manner. Further, there can be more than
25 one triple bond in the alkynyl group. The most preferred alkynyl group is the 3-propargyl-group.

Alkoxy groups preferably contain from 1 to 6 carbon atoms and are attached to the rest of the moiety via the oxygen atom. For the alkyl group contained in the alkoxy groups,
30 the same considerations apply as for alkyl groups. The most preferred alkoxy group is the methoxy group.

Aryloxy groups preferably contain from 6 to 20 carbon atoms. Those carbon atoms may be contained in one or more aromatic rings and further in side chains (for example,
35 alkyl chains) attached to the aromatic moiety. Preferred aryloxy group are the phenoxy

and the benzoxy group.

Preferred O-protecting groups in R^{14} are the aroyl groups, the acyl groups and the silyl groups. Among these most preferred is the benzoyl group.

5

Preferred silyl groups are the trialkylsilyl groups, like triethylsilyl.

Any atom in the definitions within the formulae presented herein is not limited to a specific isotope. Thus, a phosphorous atom (P) can either mean the regular ^{31}P or the radioactive ^{32}P or a mixture thereof. The same applies for hydrogen (H/D/T), carbon (C), iodine (Cl, Br, I) and nitrogen (N).

10

Preferred group $-\text{NR}^5\text{R}^6$ in the definition of R^2 , R^3 and R^4 is the $-\text{NH}_2$ group. In this case, it is evident that during chemical synthesis of compounds containing such group of formula I one of the hydrogen atoms of this amino group might be protected by suitable amino protecting group. Such protecting groups are generally known to a man skilled in the art.

15

The same applies for the definitions of R^1 .

20

During chemical synthesis, any groups $-\text{OH}$, $-\text{SH}$ and $-\text{NH}_2$ (including those groups in reporter groups) should be protected by suitable protecting groups. Further, during chemical synthesis, the compound will be attached for convenience to a solid phase. In these cases, the definitions of the substituents given above will be selected accordingly.

25

A protecting group is a chemical group that is attached to a functional moiety (for example to the oxygen in a hydroxyl group, replacing the hydrogen) to protect the functional group from reacting in an undesired way. A protecting group is further defined by the fact that it can be removed without destroying the biological activity of the molecule formed, here the binding of the nucleic acid binding compound to a nucleic acid. Suitable protecting groups are known to a man skilled in the art. Especially preferred protecting groups for example for hydroxyl groups at the 5'-end of a nucleotide or oligonucleotide are selected from the trityl group, for example dimethoxytrityl.

35

Preferred protecting groups at exocyclic amino groups in formula I are the acyl groups, most preferred the benzoyl group (Bz), phenoxyacetyl or acetyl or formyl, and the N,N-dialkylformamidine group, preferentially the dimethyl-, diisobutyl-, and the di-n-butylformamidine group.

5

The nucleic acid binding compound according to the invention preferably has a length of less than 100 subunits, more preferably of from 10 to 30 subunits. In order to be active as nucleic acid binding compound, the substituents should be chosen such that hydrogen bonds to heterocyclic groups at the nucleic acid to be bound are enabled, preferably by Watson Crick base pairing and/or in the way as disclosed in figure 1. Compounds in which the substituents do not enable such preferred hydrogen bonding, can be useful as intermediates for the preparation of nucleic acid binding compounds. Preferred nucleic acid binding compounds of the invention are those which are chemically synthesized.

15

If the nucleic acid binding compound is to be used as a probe for the determination of a nucleic acid, or any other identification of the compound or the nucleic acid is intended, any of the substituents are selected such as to contain a reporter group. While as many reporter groups can be attached as useful to label the nucleic acid compound sufficiently, it is preferred to attach only a limited number of reporter groups to a single subunit, such that recognition of nucleic acids, affinities to nucleic acids and solubility is not affected such that the probe would not be useful in hybridization assays. In a very preferred case, there will be only from 1 to 4, most preferably 1 or 2 or most preferred only one reporter group in each nucleic acid binding compound. There are formats for the nucleic acid determination which require more than one reporter group attached to the probe. An example for such formats is disclosed in WO92/02638. In this case, one of the reporter groups will be a fluorescence emitter, while the other is a fluorescence quencher.

Reporter groups are generally groups that make the nucleic acid binding compound as well as any nucleic acids bound thereof distinguishable from the remainder of the liquid, i.e. the sample (nucleic acid binding compounds having attached a reporter group can also be termed labelled nucleic acid binding compounds, labelled probes or just probes). This distinction can be either effected by selecting the reporter group from the group of directly or indirectly detectable groups or from the groups of immobilized

35

- or immobilizable groups. Directly detectable groups are for example fluorescent compounds, like fluorescein and its derivatives, like hexachlorofluorescein and hexafluorofluorescein, rhodamines, psoralenes squaraines, porphyrines, fluorescent particles, bioluminescent compounds, like acridinium esters and luminol, or the
- 5 cyanine dyes, like Cy-5. Examples of such compounds are disclosed in EP 0 680 969. Further, spin labels like TEMPO, electrochemically detectable groups, ferrocene, viologene, heavy metal chelates and electrochemiluminescent labels, like ruthenium bipyridyl complexes, and naphthoquinones, quencherdyes, like dabcyI, and nuclease active complexes, for example of Fe and Cu, are useful detectable groups. Indirectly
- 10 detectable groups are groups that can be recognized by another moiety which is directly or indirectly labelled. Examples of such indirect detectable groups are for example haptens, like digoxigenin or biotin. Digoxigenin for example can be recognized by antibodies against digoxigenin. Those antibodies may either be labelled directly or can be recognized by labelled antibodies directed against the (antidigoxigenin) antibodies.
- 15 Formats based on the recognition of digoxigenin are disclosed in EP-B-0 324 474. Biotin can be recognized by avidin and similar compounds, like streptavidin and other biotin binding compounds. Again, those compounds can be labelled directly or indirectly.
- 20 The reporter group can further be a nucleotide sequence which does not interfere with other nucleotide sequences in the sample. The sequence can therefore be specifically recognized by nucleotide containing a complementary sequence. This nucleotide sequence can be labelled directly or indirectly or can be immobilizable or immobilized.
- 25 A reporter group can further be a solid phase. Attachment of the nucleic acid binding compound with solid phase can be either directly or indirectly as pointed out above for the detectable group.
- Direct labelling can be effected by covalent coupling of a nucleic acid binding
- 30 compound to a reactive group on the solid phase, i.e. preferably via a linker. Indirect labelling can be made similar as disclosed above for the detectable groups. Preferably, indirect attachment is non-covalently by biospecific interactions, for example those selected from the group of hapten-antibody, vitamin-receptor and nucleic acid-complementary nucleic acid. Again, those interactions and their use in nucleic acid
- 35 assays is known to a man skilled in the art.

Solid phases that are useful for immobilization of the probe according to the invention are preferably selected from the group of polystyrene, polyethylene, polypropylene, glass and TiO₂. The formats of such solid phases can be selected according to the needs of the instrumentation and format of the assay. For example, a solid phase may assume the form of a bead or a vessel.

The term reporter group and the specific embodiments preferably include a linker which is used to connect the moiety intended to be used (the actual solid phase or the fluorophoric moiety, to the position of attachment as the reporter group. The linker will provide flexibility such that the nucleic acid binding compound can bind the nucleic acid sequence to be determined without major hindrance by the solid phase. Linkers, especially those that are not hydrophobic, for example based on consecutive ethylenoxy units, for example as disclosed in DE 3943522 are known to a man skilled in the art.

From the above explanation, it becomes clear that the invention would still work, even if the backbone of the probe is not an oligonucleotide in the strict sense. There were described in the last years nucleic binding compounds that have similar properties like oligonucleotides, but differ in their backbone. The backbone is generally considered to be the part of the nucleic acid binding compound that bears the bases, mostly in linear manner, bound to identical or not identical subunits. The most popular backbone is the naturally occurring sugar phosphate backbone of nucleic acids (containing either ribonucleoside subunits (RNA), deoxyribonucleoside subunits (DNA) or peptide nucleic acid subunits (PNA)). Therefore, in a preferred embodiment, the backbone comprises sugar and phosphate moieties. In a further preferred embodiment, the sugar configuration is selected from the group consisting of the α -D-, β -D-, α -L- and β -L-configurations, most preferred the compound contains at least one 2'-deoxy- β -D-erythro-pentofuranosyl moiety or one β -D-ribofuranosyl moiety.

Preferred, D is the glycosid C-1 of a sugar moiety of the compound according to the invention. Preferred compounds of formula VI are those wherein R¹ is NH₂, W is N, Z is N, Y is C, X is N and R¹⁴ is H.

Further preferred nucleic acid binding compounds contain at least one group of formula I, wherein R¹ is the group -NR²⁰R²¹, which are either 2-aza-adenosine or

derivatives thereof. As derivatives of 2-aza-adenine there are considered here compounds that provide hydrogen bonding via the same atoms as 2-aza-adenine to G and dG in a nucleic acid bound to the nucleic acid binding compound of the invention. The most preferred group of formula I is 2-aza-adenine, bound to the backbone via the N⁹ atom. Those groups both discriminate clearly between the natural nucleobases and in addition provide a very similar stability as the A-T base pair.

The nucleic acid binding compound will be constructed such that it contains a nucleobase sequence which is substantially complementary to the nucleic acid to be determined or the nucleic acid to which it is intended to be bound by base pairing. As those nucleic acids will usually contain at least once any of the naturally occurring nucleobases Ade, Cyt, Gua and Thy or Ura, the nucleic acid binding compound according to the invention will also contain any of those four bases. However, according to the invention, at least one of the heterocyclic groups in a position of the nucleic acid binding compound located vis-à-vis the G in the nucleic acid to be determined as replaced by the heterocyclic base of formula I. If there is more than one G in the sequence to which the nucleic acid binding compound is intended to be hybridized on the nucleic acid, preferably as many C's in the nucleic acid binding compound are chosen to be heterocyclic groups of formula I as necessary to provide the T_m as intended.

However, the nucleic acid binding compounds of the invention display the same base pairing selectivity also versus other heterocyclic groups in the position located vis-à-vis the position of the nucleic acid binding compound at which the group of formula I is located, especially versus derivatives of G, for example c⁷G_d, z⁸c⁷G_d and z⁸G_d. In these nominations, c⁷ means that in the 7-position there will be a carbon atom and z⁸ will correspondingly mean that in the 8-position there will be a nitrogen atom. Further, derivatives of G that can be recognized according to the invention, are nucleobases G which are labelled by the attachment of detectable groups, and iso G_d.

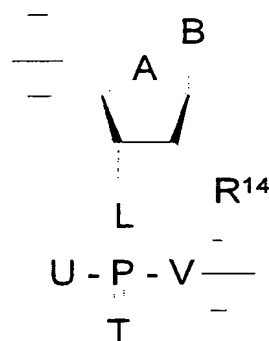
The nucleic acid can also contain a heterocyclic group of formula I itself. The corresponding base on the nucleic acid binding compound will preferably be selected such as to base pair with this group, for example to be dG. The nucleic acid can contain natural and/or non-natural bases, for example 7-deaza-dGTP. Thus, the term nucleic acid will be construed in the present invention very broadly. Nucleic acids having mixed

base sequences being preferred.

The nucleic acid binding compound according to the invention will bind to nucleic acids preferably in the antiparallel mode. However, by carefully selecting the nucleobases of a nucleic acid and/or of the nucleic binding compound, the binding can also be forced to be in the parallel mode. Parallel hybridization of nucleic acids containing iso-C and iso-G are for example disclosed in EP 0 624 161.

Preferred nucleic acid binding compounds are those, wherein the backbone comprises one or more moieties of the general formula II

Formula II



wherein

- A is selected from the group consisting of O, S and N-(C₁-C₁₀)-alkyl,
- 15 L is selected from the group consisting of oxy, sulfanediyl and -NR²²-,
- T is selected from the group consisting of oxo, thioxo and selenoxo,
- U is selected from the group consisting of -OH, -O-reporter group, -SH, -S reporter group -SeH, -(C₁-C₁₀)-alkoxy, (C₁-C₁₀)-alkyl, -(C₆-C₂₂)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -NR²³R²⁴, and
- 20 -O-(C₁-C₁₀)-alkyl-O-(C₁-C₁₀)-alkyl-R²⁵, or wherein -NR²³R²⁴ can together with N be a 5-6-membered heterocyclic ring,
- V is selected from the group consisting of oxy, sulfanediyl or -NR²²-,
- R¹⁴ is selected from the group consisting of -H, -OH, -(C₁-C₁₀)-alkoxy, -(C₂-C₁₀)-alkenyloxy, -halogen, -azido, -O-allyl, -O-alkinyl, and -NH₂,
- 25 R²² is independently selected from the group of -H and -(C₁-C₁₀)-alkyl,

R^{23} and R^{24} are independently selected from the group consisting of $-(C_1-C_{10})$ -alkyl, $-(C_1-C_{20})$ -aryl, $-(C_6-C_{14})$ -aryl- $-(C_1-C_{10})$ -alkyl, $-(C_1-C_6)$ -alkyl- $[NH(CH_2)_c]_d-NR^{26}R^{27}$ and a reporter group,

R^{25} is selected from the group consisting of $-H$, $-OH$, $-halogen$, $-amino$, $-(C_1-C_{18})$ -alkylamino, $-COOH$, $-CONH_2$ and $-COO(C_1-C_4)$ -alkyl and a reporter group,

R^{26} and R^{27} are independently selected from the group consisting from $-H$, $-(C_1-C_6)$ -alkyl, and $-(C_1-C_4)$ -alkoxy- $-(C_1-C_6)$ -alkyl and a reporter group,

c is an integer from 2 to 6,

d is an integer from 0 to 6, and

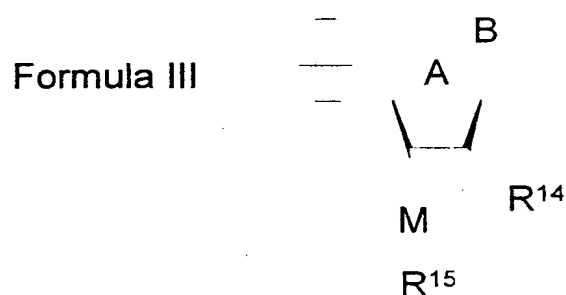
B is a moiety of formula I,

wherein any alkyl, alkenyl and alkynyl can be substituted or unsubstituted,

and any salts thereof.

The preferred definitions of the groups as defined under formula I apply to formula II and the following formulae, if not indicated otherwise.

A preferred subject of the invention is therefore a nucleic acid binding compound as outlined above, wherein the backbone comprises one or more moieties of the general formula III



wherein

A is selected from the group consisting of O , S and N - (C_1-C_6) -alkyl,

- M is selected from the group consisting of oxy, sulfanediyl, $-NR^{22}$ -, $-(C_1-C_{10})$ -alkyl-, or $-O-(C_1-C_{10})$ -alkyl-O-, and $-S-(C_1-C_{10})$ -alkyl-O- and $-NR^{22}-(C_1-C_6)$ -alkyl-O-,
- 5 R^{22} is selected from the group of $-H$ -, $-(C_1-C_{10})$ -alkyl, a protecting group and a reporter group,
- R^{14} is selected from the group consisting of $-H$ -, $-OH$ -, $-(C_1-C_{10})$ -alkoxy, $-(C_2-C_{10})$ -alkenyloxy, $-(C_2-C_{10})$ -alkynyloxy, -halogen, -azido, SH -, $-(C_1-C_{10})$ -alkylmercapto and $-NH_2$,
- 10 R^{15} is selected from the group consisting of $-H$ -, $-(C_1-C_6)$ -alkyl, $-(C_2-C_{10})$ -alkenyl, $-(C_2-C_{10})$ -alkynyl, $-(C_2-C_{10})$ -alkyl-carbonyl, $-(C_3-C_{19})$ -alkenyl-carbonyl, $-(C_3-C_{19})$ -alkynyl-carbonyl, $-(C_6-C_{14})$ -aryl- $-(C_1-C_{10})$ -alkyl, a solid phase and a group of formula IV

Formula IV



$$T$$

wherein

15

- T is selected from the group consisting of oxo, thioxo and selenoxo, and
- U is selected from the group consisting of $-OH$ -, $-O$ -reporter group, $-SH$ -, $-SeH$ -, $-(C_1-C_{10})$ -alkoxy, $-(C_1-C_{10})$ -alkyl, $-(C_6-C_{22})$ -aryl, $-(C_6-C_{14})$ -aryl- $-(C_1-C_{10})$ -alkyl, $-NR^{23}R^{24}$, and $-O-(C_1-C_{10})$ -alkyl-O- $-(C_1-C_{10})$ -alkyl- R^{25} , or wherein
- 20 $NR^{23}R^{24}$ can together with N be a 5-6-membered heterocyclic ring,
- R^{23} and R^{24} are independently selected from the group consisting of $-(C_1-C_{10})$ -alkyl, $-(C_1-C_{20})$ -aryl, $-(C_6-C_{14})$ -aryl- $-(C_1-C_{10})$ -alkyl, $-(C_1-C_6)$ -alkyl- $[NH(CH_2)_c]_d-NR^{26}R^{27}$,
- R^{25} is selected from the group consisting of $-H$ -, $-OH$ -, -halogen, -amino,
- 25 $-(C_1-C_{18})$ -alkylamino, $-COOH$ -, $-CONH_2$ and $-COO(C_1-C_4)$ -alkyl,
- R^{26} and R^{27} are independently selected from the group consisting from $-H$ -, $-(C_1-C_6)$ -alkyl, and $-(C_1-C_4)$ -alkoxy- $-(C_1-C_6)$ -alkyl
- R^{29} is selected from the group consisting of $-OR^{30}$ and $-SR^{30}$,

R^{30} is selected from the group consisting of -H, $-(C_1-C_{10})$ -alkyl, $-(C_2-C_{10})$ -alkenyl, $-(C_6-C_{22})$ -aryl, a protecting group, a solid phase and a reporter group

B is the link to a moiety of formula I,

5

and any salts thereof.

For the definitions and preferences the particulars apply as outlined for the substituents under formulae I and II, if not specified otherwise specifically for formula III.

10

Preferably, in compounds of formula II, R^{14} is hydrogen. Preferred definition of L is oxy. Preferred definition of U is -OH and -O-reporter group. Preferred definition of V is oxy. Preferred definition of c is an integer from 2 to 4, and of d an integer from 0 to 2.

15

Compounds of formula II are especially suited to contain the heterocyclic moiety of the invention as an integrated part (preferably not at one of the termini) of the nucleic acid binding compound.

20 The group $NR^{23}R^{24}$ is preferably selected from the group consisting of dialkylamino groups. In case of this group together with the forming of 5- or 6-membered heterocyclic ring, it assumes preferably the definition of pyrrolidinyl or piperidinyl.

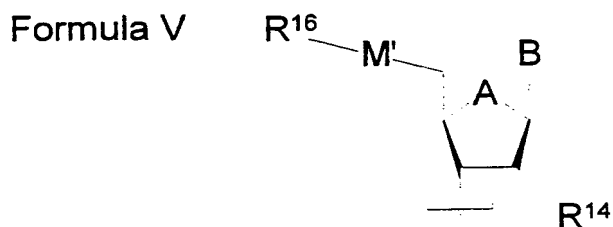
Preferred aryl group is the phenyl or naphthyl moiety, either unsubstituted or substituted by one or more of amino, -aminoalkyl, $-O-(C_1-C_{10})$ -alkyl, $-S-(C_1-C_{10})$ -alkyl, $-(C_1-C_{10})$ -alkyl, sulfonyl, sulfenyl, sulfinyl, nitro and nitroso. Most preferred aryl group is the phenyl group. Preferred arylalkyl group is the benzyl group. The preferred alkylamino group is the ethylamino group. The preferred $-COO(C_1-C_4)$ alkyl group contains one or two carbon atoms in the alkyl moiety (methyl or ethyl esters).

30

Nucleic acid binding compounds, wherein the group of formula I is attached to submit, for example the nucleotide, at the 3'-terminus of the compound, are useful either as starting compound for longer compounds or/and as end-labelled probes. This group of compounds is especially preferred because the terminal position of probes generally is the most tolerant in view of attachment of chemical moieties.

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A preferred subject of the invention is a nucleic acid binding compound as outlined above comprising a backbone moiety of the formula V

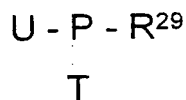


5

wherein

- A is selected from the group consisting of O, S and N-(C₁-C₆)-alkyl,
 M' is selected from the group consisting of oxy, sulfanediyl, -NR²²-, -(C₁-C₁₀)-
 10 alkyl, or -O-(C₁-C₁₀)-alkyl-O-, and -S-(C₁-C₁₀)-alkyl-O- and -NR²²-(C₁-
 C₆)-alkyl-O-,
 R²² is selected from the group of -H, a protecting group, a reporter group and
 -(C₁-C₁₀)-alkyl,
 R¹⁴ is selected from the group consisting of -H, -OH, -(C₁-C₁₀)-alkoxy,
 15 -(C₂-C₁₀)-alkenyloxy, -(C₂-C₁₀)-alkynyloxy, -halogen, azido, -SH, -S-(C₁-
 C₆)-alkylmercapto and NH₂,
 R¹⁶ is selected from the group consisting of -H, -(C₁-C₈)-alkyl, -(C₂-C₁₈)-
 alkenyl, -(C₂-C₁₈)-alkynyl, -(C₂-C₁₈)-alkyl-carbonyl, -(C₃-C₁₉)-alkenyl-
 carbonyl, -(C₃-C₁₉)-alkynyl-carbonyl, -(C₆-C₁₄)-aryl-(C₁-C₈)-alkyl, a
 20 protective group or a compound of formula IV

Formula IV



wherein

T is selected from the group consisting of oxo, thioxo and selenoxo,

- U is selected from the group consisting of -OH, -SH, -SeH, -(C₁-C₁₀)-alkoxy, -(C₁-C₁₀)-alkyl, -(C₆-C₂₂)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -NR²³R²⁴, and -O-(C₁-C₁₀)-alkyl-O-(C₁-C₁₀)-alkyl-R²⁵, wherein NR²³R²⁴ can together with N be a 5-6-membered heterocyclic ring,
- 5 R²³ and R²⁴ are independently selected from the group consisting of -(C₁-C₁₀)-alkyl, -(C₁-C₂₀)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -(C₁-C₆)-alkyl-[NH(CH₂)_c]_d-NR²⁶R²⁷,
- R²⁵ is selected from the group consisting of -H, -OH, -halogen, -amino, -(C₁-C₁₈)-alkylamino, -COOH, -CONH₂ and -COO(C₁-C₄)-alkyl,
- 10 R²⁶ and R²⁷ are independently selected from the group consisting from -H, -(C₁-C₆)-alkyl, and -(C₁-C₄)-alkoxy-(C₁-C₆)-alkyl
- R²⁹ is selected from the group consisting of -OR³⁰ and -SR³⁰,
- R³⁰ is selected from the group consisting of -H, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₆-C₂₂)-aryl, a protecting group, a solid phase and a reporter
- 15 group, and
- B is the link to a moiety of formula I,

wherein any alkyl, alkenyl and alkynyl can be substituted or unsubstituted,

- 20 and any salts thereof.

Those compounds are compounds that can be used as 5'-terminally labelled probes. Regarding the definitions of the substituents, the definitions as given above apply if not indicated otherwise.

25

A very preferred compound is a compound of formula V, wherein M is O, R¹⁶ is H and R¹⁴ is selected from the group consisting of hydrogen and hydroxyl.

- The backbone of the nucleic acid binding compound has the function to bear the base pairing heterocycles such that the compound can bind to a nucleic acid having a complementary sequence. Preferably, the degree of complementarity in the naturally occurring bases will be in the range from 70 % up to 100 % in a stretch of bases in a region effecting binding, compared to the stretch of same length in the region of the nucleic acid to be bound. Deletions and insertions of subunits in each sequence will
- 30 therefor, in this calculation, be counted as gaps until the next fitting base and thus
- 35

reduce complementarity by as many bases as the gap contains.

Preferred backbone contains sugar-phosphate moieties. From these, deoxy sugar containing backbones are further preferred.

5

Each moiety in the backbone bearing a moiety capable of base pairing to a nucleic acid of complementary sequence, including the moieties of the invention, are termed a subunit. Compounds are known that have backbones mixed of different kinds of subunits. Recently, a new kind of non-natural nucleic acid binding compounds was described. They are termed Peptide Nucleic Acids (PNA), as they contain at least one peptide bond between the subunits (WO 92/20702). The nucleic acid binding compound of the present invention can have any length. However, due to the convenience of chemical synthesis, compounds of a length of less than 100, more preferably from 10 to 30 subunits, for example nucleosides, are preferred.

10

The nucleic acid binding compound of the present invention can be prepared analogous to known methods.

15

In a first option which is particularly suitable for short compounds, the compounds are produced by chemical synthesis (multistep oligomerisation) using chemically activated derivatives of the subunits (monomers), at least one of them containing the modified base of the invention. Preferably, reactive groups in the monomers that are not involved in the actual reaction step of the oligomerisation reaction are protected using an appropriate protecting group. Such protecting groups are well known in the art and there will be no major change in protection and synthesis strategy if the groups of the invention are used.

20

An activated subunit is a subunit containing a substituent especially suited for chemical reaction with a predetermined substituent in another subunit, on the surface of a solid phase or in an oligomer formed of subunits. Such especially suited subunits are preferably selected from the group consisting of phosphoramidites, phosphonates (like methylphosphonates), phosphotriesters, phosphothioates, phosphodithioates, boranophosphates (see Chem. Commun. 1999, 1517-1518), phosphate methyl esters, phenylphosphonates and phosphate ethyl esters.

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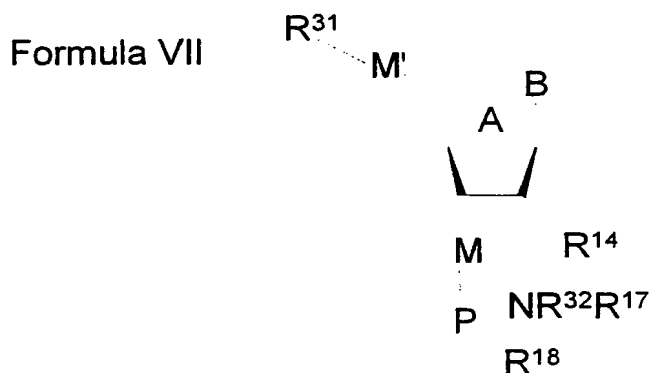
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The predetermined substituents are preferably selected from the group of $-NH_2$, $-SH$ and $-OH$.

A further subject of the invention is therefore a method for the chemical synthesis of
 5 a compound of any of claims 1 to 13 using activated subunits, wherein said subunit
 contains at least one group of formula I. There are several approaches known for the
 chemical synthesis, such as the phosphotriester method of Narang et al., Meth.
 Enzymol. 68, 90-99 (1979); the phosphodiester method of Brown et al., Meth. Enzymol.
68, 109-151 (1979); the diethylphosphoramidite method of Beaucage et al., Tetrahedron
 10 Lett. 22, 1859-1862 (1981); and the solid support method described in the U.S. Patent
 Specification No. 4,458,066 and in Methods in Molecular Biology, Ed. S. Agrawal, Vol.
 20, Humana Press, Totowa, NJ, 1993. The most preferred method of chemical synthesis
 uses the phosphoramidite approach. A particularly preferred method uses a activated
 subunit one or more compounds of general formula VII. This method has the
 15 advantage that it is very convenient and the reagents necessary, for example a
 phosphoramidite containing a group of formula I, is possible to be included easily.

A further subject of the invention are therefore compounds of the general formula VII



wherein

A is selected from the group consisting of O, S and N-(C₁-C₆)-alkyl,
 M and M' are independently selected from the group consisting of oxy,
 25 sulfanediyl, $-NR^{22}$, $-(C_1-C_{10})$ -alkyl, or $-O-(C_1-C_{10})$ -alkyl-O-, and $-S-$
 (C_1-C_{10}) -alkyl-O- and $-NR^{22}-(C_1-C_6)$ -alkyl-O-,
 R²² is selected from the group of $-H$ and $-(C_1-C_{10})$ -alkyl,

R^{14} is selected from the group consisting of -H, -OR³¹, -(C₁-C₁₀)-alkoxy, -(C₂-C₁₀)-alkenyloxy, -(C₂-C₁₀)-alkynyloxy, -halogen, -azido NHR³¹, SR³¹ and -NH₂,

R^{31} is a protecting group or a reporter group,

5 R^{32} and R^{17} are independently selected from the group consisting of -H, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl and -(C₆-C₂₂)-aryl,

R^{18} is selected from the group consisting of substituted or unsubstituted -(C₁-C₆)-alkyl, unsubstituted -(C₁-C₆)-alkoxy or -(C₁-C₆)-alkoxy substituted one or more times by a group selected from the group consisting of -halogen, p-nitroaryloxy and -cyano, and

10

B is a group of formula I.

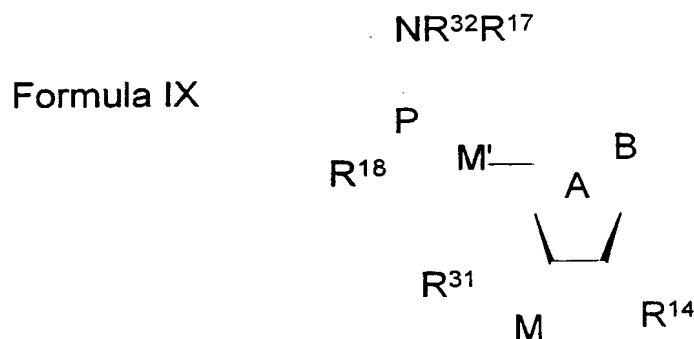
Preferred compounds of formula VII are those wherein the group of formula I is not 2-aza-hypoxanthine. In a preferred embodiment, the group of formula I in

15

formula VII contains at least one reporter group. Most preferable, the group of formula I contains exactly one reporter group.

Most preferred in such compounds, in -NR⁵R⁶ at least one of R⁵ and R⁶ is a protecting group.

20 Further subject of the invention are compounds of general formula IX



wherein

25 A is selected from the group consisting of O, S and N-(C₁-C₆)-alkyl,

M and M' are independently selected from the group consisting of oxy, sulfanediyl, -NR²², -(C₁-C₁₀)-alkyl, or -O-(C₁-C₁₀)-alkyl-O-, and -S-(C₁-C₁₀)-alkyl-O- and -NR²²-(C₁-C₆)-alkyl-O-,

R²² is selected from the group of -H and -(C₁-C₁₀)-alkyl,

5 R¹⁴ is selected from the group consisting of -H, -OR³¹, -(C₁-C₁₀)-alkoxy, -(C₂-C₁₀)-alkenyloxy, -(C₂-C₁₀)-alkynyloxy, -halogen, -azido NHR³¹, SR³¹ and -NH₂,

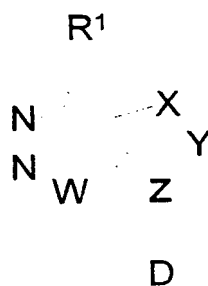
R³¹ is a protecting group or a reporter group,

10 R³² and R¹⁷ are independently selected from the group consisting of -H, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl and -(C₆-C₂₂)-aryl,

R¹⁸ is selected from the group consisting of substituted or unsubstituted -(C₁-C₆)-alkyl, unsubstituted -(C₁-C₆)-alkoxy or -(C₁-C₆)-alkoxy substituted one or more times by a group selected from the group consisting of -halogen, p-nitroaryloxy and -cyano, and

15 B is a group of formula I

Formula I



20 wherein

W is selected independently from X, Y and Z from the group consisting of N and CR²,

Z is selected from the group consisting of N and C with the proviso that

25

- if Z is N, then

X independently from W and Y is selected from the group consisting of N and CR^3 , and

Y independently from W and X is selected from the group consisting of N and CR^4 ,

5 and the bond between X and Y is a double bond and the bond between Y and Z is a single bond, and

- if Z is C, then

10 X is NR^{33} , and

Y is selected from the group consisting of N and CR^4 and the bond between Z and Y is a double bond and the bond between X and Y is a single bond,

15 R^1 , R^2 , R^3 and R^4 are independently selected from the group consisting of -H, -halogen, $-\text{OR}^{13}$, $-\text{SR}^{19}$, $-(\text{C}_1-\text{C}_{10})$ -alkyl, $-(\text{C}_2-\text{C}_{10})$ -alkenyl, $-(\text{C}_2-\text{C}_{10})$ -alkynyl, $-\text{NO}_2$, $-\text{NR}^5\text{R}^6$, -cyano, and $-\text{C}(=\text{O})\text{R}^{11}$,

R^{11} is selected from the group consisting of -OH, $-(\text{C}_1-\text{C}_6)$ -alkoxy, $-(\text{C}_6-\text{C}_{22})$ -aryloxy, and NHR^{12} ,

20 R^5 , R^6 , R^{12} , R^{13} , R^{19} and R^{33} are selected independently from the group consisting of -H, $-(\text{C}_1-\text{C}_{10})$ -alkyl, $-(\text{C}_2-\text{C}_{10})$ -alkenyl, $-(\text{C}_2-\text{C}_{10})$ -alkynyl, $-(\text{C}_6-\text{C}_{22})$ -aryl, a protecting group and a reporter group,

r and s are independently of each other an integer of 1 to 18,

25 D is the position of attachment of the group to the rest of the nucleic acid binding compound, and

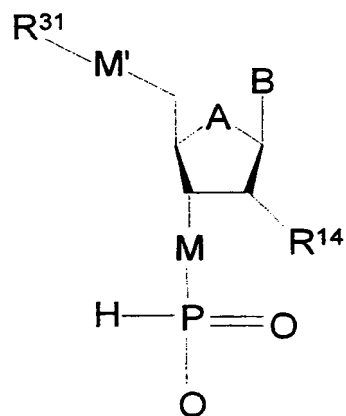
alkyl, alkenyl and alkynyl being unsubstituted or substituted by one or more moieties selected from the group consisting of -halogen, $-\text{S}-(\text{C}_1-\text{C}_6)$ -alkyl, $-(\text{C}_1-\text{C}_6)$ -alkoxy, $-\text{NR}^5\text{R}^6$, $-\text{CO}-\text{R}^{11}$, $-\text{NH}-\text{CO}-\text{NR}^5\text{R}^6$, $-\text{NH}-\text{CSNR}^5\text{R}^6$ and $-\text{[O}-(\text{CH}_2)_r]_s-\text{NR}^5\text{R}^6$,

30 with the proviso that at least one of R^5 and R^6 of $-\text{NR}^5\text{R}^6$ is a protecting group.

Those compounds can be used like those of formula VII in chemical synthesis.

A further subject of the invention are compounds of general formula X

Formula X



wherein

M and M' are independently selected from the group consisting of oxy,
 5 sulfanediyl, -NR²², -(C₁-C₁₀)-alkyl, or -O-(C₁-C₁₀)-alkyl-O-, and -S-
 (C₁-C₁₀)-alkyl-O- and -NR²²-(C₁-C₆)-alkyl-O-,

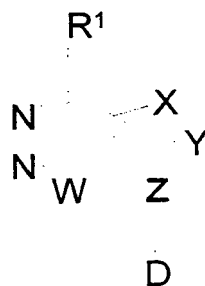
R²² is selected from the group of -H and -(C₁-C₁₀)-alkyl,

R¹⁴ is selected from the group consisting of -H, -OR³¹, -(C₁-C₁₀)-alkoxy,
 10 -(C₂-C₁₀)-alkenyloxy, -(C₂-C₁₀)-alkynyloxy, -halogen, -azido NHR³¹, SR³¹
 and -NH₂,

R³¹ is a protecting group or a reporter group,

B is a group of formula I

Formula I



15

wherein

W is selected independently from X, Y and Z from the group consisting of N
 and CR²,

Z is selected from the group consisting of N and C with the proviso that

- if Z is N, then

5 X independently from W and Y is selected from the group
consisting of N and CR³, and
Y independently from W and X is selected from the group
consisting of N and CR⁴,
and the bond between X and Y is a double bond and the bond
10 between Y and Z is a single bond, and

- if Z is C, then

15 X is NR³³, and
Y is selected from the group consisting of N and CR⁴ and
the bond between Z and Y is a double bond and the bond between X
and Y is a single bond,

20 R¹, R², R³ and R⁴ are independently selected from the group consisting of -H, -
halogen, -OR¹³, -SR¹⁹, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₂-C₁₀)-alkynyl,
-NO₂, -NR⁵R⁶, -cyano, and -C(=O)R¹¹,

R¹¹ is selected from the group consisting of -OH, -(C₁-C₆)-alkoxy, -(C₆-C₂₂)-
aryloxy, and NHR¹²,

25 R⁵, R⁶, R¹², R¹³, R¹⁹ and R³³ are selected independently from the group consisting
of -H, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₂-C₁₀)-alkynyl, -(C₆-C₂₂)-aryl, a
protecting group and a reporter group,

r and s are independently of each other an integer of 1 to 18,

D is the position of attachment of the group to the rest of the nucleic acid
binding compound, and

30 alkyl, alkenyl and alkynyl being unsubstituted or substituted by one or more
moieties selected from the group consisting of -halogen, -S-(C₁-C₆)-alkyl, -(C₁-
C₆)-alkoxy, -NR⁵R⁶, -CO-R¹¹, -NH-CO-NR⁵R⁶, -NH-CSNR⁵R⁶ and -[O-
(CH₂)_i]_s-NR⁵R⁶.

35 Those compounds are also useful in chemical synthesis.

In another option which is more suited for long oligomers and those based on natural backbones, the oligomers are produced enzymatically. In this case, a starting oligomer is reacted with a polymerase and a triphosphate or modified triphosphate such that a monophosphate or a modified monophosphate is attached to a terminus of the oligomer, thus elongating the oligomer. Also for this method, the man skilled in the art will know several possible formats, like the nick-translation approach, or the simple primer extension (J. Sambrook, E.F. Fritsch, T. Maniatis, Molecular Cloning - A laboratory Manual, Cold Spring Harbor Laboratory Press 1989).

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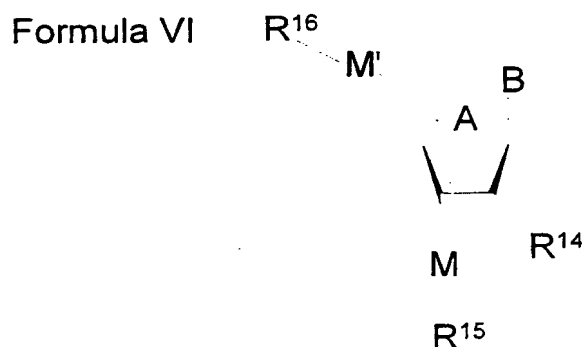
For example, the incorporation of z^2A_d into a DNA sequence can be performed via conventional methods, e.g. by polymerase-catalyzed incorporation of z^2A_d -5'-triphosphate (11).

15

A further subject of the invention is therefore a method for the enzymatic synthesis of a nucleic acid binding compound according to the invention comprising reacting a triphosphate subunit with a primer using a nucleic acid as a template for the elongation of the primer, wherein the triphosphate subunit contains a heterocyclic group of formula I. Preferably, the triphosphate subunit has the formula VI.

20

A further subject of the present invention are therefore compounds of the general formula VI



wherein

25

A is selected from the group consisting of O, S and N-(C_1 - C_6)-alkyl,

R^{14} is selected from the group consisting of -H, -OH, $-(C_1-C_{10})$ -alkoxy, O-protecting group, S-protecting group, NH-protecting group, $-(C_2-C_{10})$ -alkenyloxy, -halogen, -azido, -SH, $-(C_1-C_6)$ -alkylmercapto and $-NH_2$,

R^{15} and R^{16} are independently selected from the group consisting of -H, $-(C_1-C_8)$ -alkyl, $-(C_2-C_{18})$ -alkenyl, $-(C_2-C_{18})$ -alkynyl, $-(C_2-C_{18})$ -alkyl-carbonyl, $-(C_3-C_{19})$ -alkenyl-carbonyl, $-(C_3-C_{19})$ -alkynyl-carbonyl, $-(C_6-C_{14})$ -aryl- (C_1-C_8) -alkyl, a protecting group or a compound of formula IV

Formula IV

U - P - R^{29}

T

wherein

T is selected from the group consisting of oxo, thioxo and selenoxo,

U is selected from the group consisting of -OH, -SH, -SeH, $-(C_1-C_{10})$ -alkoxy, $-(C_1-C_{10})$ -alkyl, $-(C_6-C_{22})$ -aryl, $-(C_6-C_{14})$ -aryl- (C_1-C_{10}) -alkyl, $-NR^{23}R^{24}$, and $-O-(C_1-C_{10})$ -alkyl- $O-(C_1-C_{10})$ -alkyl- R^{25} , or wherein $NR^{23}R^{24}$ can together with N be a 5-6-membered heterocyclic ring,

R^{23} and R^{24} are independently selected from the group consisting of $-(C_1-C_{10})$ -alkyl, $-(C_1-C_{20})$ -aryl, $-(C_6-C_{14})$ -aryl- (C_1-C_{10}) -alkyl, $-(C_1-C_6)$ -alkyl- $[NH(CH_2)_C]_d-NR^{26}R^{27}$,

R^{25} is selected from the group consisting of -H, -OH, -halogen, amino, $-(C_1-C_{18})$ -alkylamino, -COOH, -CONH₂ and COO (C_1-C_4) -alkyl,

R^{26} and R^{27} are independently selected from the group consisting from -H, $-(C_1-C_6)$ -alkyl, and $-(C_1-C_4)$ -alkoxy- (C_1-C_6) -alkyl,

R^{29} is selected from the group consisting of $-OR^{30}$ and $-SR^{30}$,

R^{30} is selected from the group consisting of -H, $-(C_1-C_{10})$ -alkyl, $-(C_2-C_{10})$ -alkenyl, $-(C_6-C_{22})$ -aryl, a protecting group, a diphosphate and a reporter group, and

M and M' are independently selected from the group consisting of oxy, sulfanediyl, $-NR^{22}$, $-(C_1-C_{10})$ -alkyl, or $-O-(C_1-C_{10})$ -alkyl-O-, and $-S-(C_1-C_{10})$ -alkyl-O- and $-NR^{22}-(C_1-C_6)$ -alkyl-O-,

R^{22} is selected from the group of -H and $-(C_1-C_{10})$ -alkyl, and

B is a moiety of formula I,

wherein any alkyl, alkenyl and alkynyl can be substituted or unsubstituted, and wherein at least one of R^{15} and R^{16} is not a group of formula IV with the proviso that

MR^{16} , MR^{15} and R^{14} are not each -OH if R^1 is $-NH_2$ and if either

- W and X and Y and Z is N, or
- W and X and Z is N and Y is CR^4 , or
- W and Y and Z is N and X is CR^3 .

10

Most preferred in these compounds - MR^{16} is a triphosphate group and - MR^{15} is OH. The most preferred compound is the one in which R^{14} is -OH.

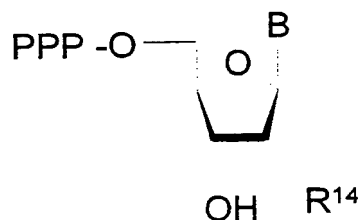
Those compounds are especially the compounds, wherein the heterocyclic moiety of the invention is contained not at the terminal position of the nucleic acid binding compound.

Preferred compounds are those, wherein M is oxy or sulfanediyl, R^{16} is a compound of formula IV wherein U is -OH, T is oxo or thioxo, R^{29} is $-OR^{30}$ and R^{30} is a disphosphate group and the salts thereof.

20

Most preferred compounds are of formula VIII

Formula VIII



25

wherein

PPP is a triphosphate group,

R^{14} is selected from the group consisting of -H, -OH, $-(C_1-C_{10})$ -alkoxy, $-(C_2-C_{10})$ -alkenyloxy, $-(C_2-C_{10})$ -alkynyloxy halogen, -azido and NH_2 , and

B is a group of formula I,
with the proviso that R^{14} is not OH if B is 2-azaadenine.

In another preferred embodiment, the group of formula I in compounds of formula VII
5 is selected from the group consisting of groups of formula I, wherein either

- W is N, Z is N, Y is N and X is CR^3 , or
- W is N, Z is C, Y is N and X is CR^3 , or
- W is N, Z is N, Y is N and X is N.

10

A further subject of the invention is a method for the enzymatic synthesis of a nucleic acid binding compound of the invention comprising reacting a triphosphate subunit with a primer using a nucleic acid template for the elongation of the primer, wherein the triphosphate subunit contains a heterocyclic group of formula I.

15

More preferable, this method uses as a triphosphate subunit a compound of formula VIII as defined above.

The compounds of general formulae VI, VII and VIII can be prepared from compounds
20 readily available. In a first embodiment, the compounds of formula VI characterized by the fact that the atom in the 2-position (purine numbering) are prepared by choosing the corresponding compound having a carbon atom in this position (preferably wherein MR^{15} is OH and $M'R^{16}$ is OH and R^1 is NH_2 , reacting it with 2-chloroacetaldehyde, cleaving the pyrimidine ring using alkaline conditions and again closing the ring using
25 sodium nitrite and thereafter hydrolyzing the imidazole ring introduced by the aldehyde. This reaction yields a replacement of the carbon atom of the 2-position by a nitrogen atom.

This compound can then be reacted with reagents to attach protecting groups or/and
30 activating groups, like phosphoramidites or phosphates, to the hydroxyl groups intended to be used in oligomerization. Methods for attaching protecting groups, like the dimethoxytrityl group are generally known to a man skilled in the art. Also the attachment of mono-, di- and triphosphate groups are known in the art.

35 In a preferred embodiment, the method for the preparation of a compound of formula

VII, wherein R^1 is a protected amino group, $M'R^{31}$ is O-protecting group and $MPR^{18}NR^{32}R^{17}$ is O-phosphoramidite group, a compound of formula VI, wherein $M'R^{16}$ and MR^{15} are each OH and R^1 is NH_2 is subjected to conditions to protect the amino group, preferably by the protecting group dialkylaminomethylidene, then reacted with reagents to protect the 5'-hydroxyl group and then reacted with an activated phosphane to produce the phosphoramidite. The resulting compound of formula VII can be used in the chemical synthesis for the introduction of the 2-aza-purine directly. The protecting group at R^1 will be removed during the chemical synthesis.

For an alternative synthesis of 2-aza-2'-deoxyadenosine see N. Yamaji, M. Kato, *Chemistry Lett.* 1975, 311-314).

By the above methods, it is principally possible to introduce only one monomer containing the moiety of the invention, but also more than one, as the case may be.

The highest reduction of T_m will occur, if all C in the binding region are replaced. This allows fine tuning of the T_m . Of course there may also be remaining Cs in any regions that are not intended to base pair with the nucleic acid to be determined.

Compounds of formula I bis VIII, in which any of substituents R^1 , R^2 , R^3 and R^4 are selected from the group consisting of halogen, especially Cl, cyano or SR^{19} are useful as intermediate products for the synthesis of compounds wherein those substituents are selected from the group of $-NR^5R^6$ and $-OR^{13}$, preferably $-NR^5R^6$. The intermediate products can be converted into the final products by substitution reaction.

A further subject of the invention is a method for the determination of a nucleic acid comprising the steps

- providing a sample suspected to contain said nucleic acid,
- providing a nucleic acid binding compound of claim 1, which is essentially complementary to a part or all of said nucleic acid,
- contacting said sample with said nucleic acid binding compound under conditions for binding said nucleic acid binding compound to said nucleic acid,
- determining the binding product formed from said nucleic acid and said nucleic acid binding compound as a measure of the presence of said nucleic acid.

Methods for the determination of nucleic acids by hybridization are generally known in the art, for example from Sambrook et al. (cited above). They can easily be adopted to the use of the probes of the present invention. Preferably, the nucleic acid binding compound will be bound to the nucleic acid in solution, as the reaction is faster than on a solid phase. It is apparent to a man skilled in the art how to determine the T_m of the hybrid of the nucleic acid to be determined and the probe prior to the construction of an assay and its general outset. If determined once, it should be clear to the man that he should choose similar conditions in each assay using the same analyte nucleic acid.

Such determination will start with providing a sample which is suspected to contain the nucleic acid to be determined. The sample may have been subject to steps for bringing the nucleic acids in an appropriate form, for example by lysing any cells, in which the nucleic acids may be contained and would otherwise not be possible to be determined. Further, any steps to provide the sample could contain steps to purify the nucleic acids to be determined from components of an original sample that could affect the determination. Such components could be enzymes that would, if a nucleic acid is set free from cells, would digest or degrade the nucleic acid, for example RNases. It is further preferred to also remove from the nucleic acids components of the original sample that could affect amplification of the nucleic acids.

There are several possibilities for methods for the determination of nucleic acids using the nucleic acid binding compound of the invention. In a first group, the nucleic acid binding compound is used as a detectable probe. In this case, the nucleic acid binding compound will contain a detectable reporter group. Any hybrids formed from the nucleic acid binding compound and a nucleic acid can then be determined via the detectable reporter group. This group of assays can further be divided into two groups, one being the group of homogeneous assays and the other being the heterogeneous assays. In heterogeneous assays, preferably the hybrid (binding product) will be determined when bound to a solid phase. This embodiment has the advantage that any excess of probe and other components can be removed easily from the hybrid, thus make the determination easier. The hybrid formed can be captured to a solid phase either covalently, noncovalently, specifically or unspecifically. There are several embodiments which are known to a man skilled in the art.

In the so-called homogeneous assays, the hybrid formed will not be bound to a solid

phase, but will be determined either directly or indirectly in solution. A preferred example of such assays is disclosed in PCT/US 91/05571 which is incorporated by reference here. The nucleic acid binding compound of the invention is especially useful as a probe in this assay. As the assay disclosed therein may need fine tuning of melting
5 temperatures of the primers and probes used in these assays, the present invention using the modulation of a melting temperature of the hybrid formed by the nucleic acid to be determined or the amplicates thereof and the probe is especially useful. This is especially useful since selectivity could be preserved, as the T_m is reduced by choosing a(n-x)-mer oligonucleotide instead of an n-mer oligonucleotide.

10

Therefore, a further subject of the invention is a method for the determination of the presence, absence or amount of a nucleic acid in a sample comprising the steps:

- providing primers, a first primer being essentially complementary to a first
15 binding sequence of said nucleic acid, and the second primer being essentially complementary to a binding sequence of a complement of this nucleic acid, and a probe being complementary to the nucleic acid or the complement thereof between the binding sequences of said primers, said probe being labelled at different subunits by at least two different reporter groups,
- 20 - subjecting the sample with said primers and said probe under conditions favouring extension of said primers and separating said reporter groups from each other by disintegrating the probe, and
- determining the extent of disintegration of the probe via at least one of said
reporter groups,

25

wherein at least one of said primer or/and probe are a nucleic acid binding compound as defined above. Preferably, the melting point of the primers and the probe will be selected such that the T_m 's are similar, but that the T_m of the probe is higher than those of the primers.

30

In another embodiment, the nucleic acid binding compound of the invention can be used as a immobilizable or immobilized probe for binding any nucleic acids to a solid phase. Modes to immobilize the compound are disclosed above. In an assay, it is either possible to immobilize the compound before contacting it with the sample or during
35 contacting the sample with a solid phase, or even after contacting the sample with the

solid phase. In each of these cases, the nucleic acid to be determined will be bound to the solid phase and preferably, any substituents of the sample not to be determined may be washed away from the solid phase, while the compounds and the nucleic acid will remain bound. Thereafter, the hybrid bound to the solid phase can be determined by
5 known methods, for example by using detectably labelled probe as outlined above or by direct determination of the hybrid, for example by contacting the hybrid with the intercalating dyes and measuring the change on the solid phase.

10 In another embodiment, the immobilized probe is used to isolate or purify a nucleic acid.

Another preferred embodiment uses a nucleic acid binding compound which is both bound to a solid phase and labelled by a detectable reporter group. The label in this case preferably is a group the detectable properties of which will change when the nucleic
15 acid to be determined will bind to the probe. Again, those compounds are known in the art.

The man skilled in the art will be in the position to design the sequence of a nucleic acid binding compound when knowing the sequence of the nucleic acid to which the nucleic
20 acid binding compound is intended to be bound. In almost all cases, the nucleic acid will have a sequence containing all four natural nucleic bases. In this case, it is preferable to choose all bases that when bound to the particulars stretch of the nucleic acid are located at positions capable of base pairing to C, A and T of the nucleic acid to be G, T and A respectively. However, these bases can be replaced by equivalent bases
25 base pairing to the mentioned bases in the nucleic acid. The base being at the position capable of base pairing to G in the nucleic acid, will now be replaced one or more time in the nucleic acid binding compound by a group of I. As outlined in this invention, the group of I can also base pair to other moieties, for example to force the orientation of binding from the antiparallel (regular) mode to the parallel (non-natural) orientation.

30

A further subject of the present invention is the use of 2-azapurine in a nucleic acid binding compound as a substitute for cytosine, especially for the binding of nucleic acids the bases of which are not only consisting of G.

35 A further subject of the present invention is the use of 2-azapurine in hybridization

reactions of probes with the nucleic acid as a base at the position of the probe base pairing at G in the nucleic acid.

Further subject of the present invention is a binding product of at least one nucleic acid
5 binding compound of the invention and a nucleic acid, the nucleic acid binding
compound and the nucleic acid being bound to each other by base pairing in parallel or
antiparallel orientation. The binding product can contain one molecule of the nucleic
acid and one molecule of the nucleic acid binding compound, which form a duplex, or
the binding product can contain three strands, thus being a triplex. The triplex can
10 contain either two molecules of the nucleic acid binding compound and one strand of
the nucleic acid, or can contain one nucleic acid binding compound and two molecules
of the nucleic acid. Which kind of triplex is formed, is dependent upon the
concentration and the complementarity of the nucleic acid binding compound and the
nucleic acid.

15 The present invention provides for the possibility to choose similar T_m s for a number
of probes with different sequence. Thus, a particular subject of the invention are
methods, wherein nucleic acids of different sequences are to be isolated or determined
simultaneously.

20 In a first embodiment, this method is a so-called multiplex method for the isolation or
determination of nucleic acids. In this embodiment, probes each containing a sequence
complementary to a sequence of one of the nucleic acids (for example, nucleic acids
from different viruses, like HCV, HIV and HBV) having a T_m adapted according to the
25 invention (one or more probes containing a group of formula I) are contacted with the
sample suspected to contain said nucleic acids. Depending upon the formate, the
different nucleic acids can be determined via the hybrids formed with the probes either
as a sum or independently.

30 A second embodiment is based on arrays of probes. Especially, a subject of the
invention is a method for the determination of the presence or absence of nucleic acids
each comprising a particular sequence in a sample comprising the steps

- contacting said sample with a solid phase having immobilized on its surface nucleic acid binding compounds each containing a sequence complementary to one of said particular sequences of said nucleic acids,
 - 5 - determining on said solid phase the formation of hybrids containing a nucleic acid with a particular sequence and the nucleic acid binding compound containing the complementary sequence
- characterized in that said at least one of said nucleic acid binding compounds is a
10 compound comprising a backbone, said backbone having attached heterocyclic groups capable of base pairing to natural nucleobases at least one of said heterocyclic groups being one of the naturally occurring nucleobases characterized in that at least one other of said heterocyclic groups is a group of the general formula I.
- 15 Methods of this type are either useful when simultaneously testing a sample for the presence or absence of nucleic acids, for example a panel of different bacterial species. In this way, it is not only possible to test for one species after each other batchwise, but to receive sequence information on different nucleic acids simultaneously. Those types
20 of methods are generally known in the art. The present invention, however, has found that the nucleic acid binding compounds of the invention are particularly useful in such kind of assays, as the melting temperature can be adjusted to be very similar for nucleic acid binding compounds of difference sequence or/and length. It is evident that in order to achieve similar melting temperatures of each of the nucleic acid binding compounds,
25 not all of these different compounds need to be modified according to the invention, but the man skilled in the art will modify just those compounds which do not behave like the others, for example by having a much higher melting temperature than the melting temperatures of the other compounds on the same solid phase. With each C replaced according to the invention, the T_m may decrease by between 3 and 6 °C,
30 preferably by between 4 and 5 °C. In this respect, the man skilled in the art will, compared to the chip technology presently known, has much more flexibility to select the particular sequences used for binding the different nucleic acids. For example, the high G-C content of such regions in nucleic acids to be determined up to now prevented these regions from being useful as target regions for such assays based on
35 probes complementary to these regions. According to the present invention, even these

regions, which may be highly specific to the nucleic acid to be determined, can now be used in such assays.

Assays using array technology are generally known to a man skilled in the art, for example, from EP-A-0 476 014. Those solid phases are preferably flat carriers. On their surface, separated by pure surface, arrays, each of the arrays having bound a probe of different sequence directed to a specific particular sequence in a nucleic acid. Depending upon the needs of the actual assay, the sequence of the probe will be essentially complementary to one (or more) of the particular sequences contained in said nucleic acid to be determined. During the assay, each nucleic acid will find the probe on the surface to which it can bind. The determination of the formation of hybrids in each array on the solid phase allows the determination of the presence or absence of nucleic acids containing the particular sequence based on the change of a property in this specific array. Preferably, the evaluation of these changes will be made assisted by a computer programme, which knows in which array which particular sequence is present. One chip can contain from 2 to thousands, or even millions of arrays, each having bound a nucleic acid binding compound having a specific sequence which may be unique. However, it is even possible to determine the presence of a group of different nucleic acids, each having a sequence in common, just by using a nucleic acid binding compound being either relatively unspecific and thus binding different sequences, or by selecting the sequence of the nucleic acid binding compounds such that it is directed to a sequence which is present on different nucleic acids.

In a second, slightly different approach, such arrays can be used to determine the sequence of a nucleic acid by following the so-called "sequencing by hybridization" approach. In this mode, the sample preferably will contain mostly nucleic acids of the same sequence. This can be achieved by isolating the sequence from a mixture in which they were contained or by enriching them in-vitro or in-vivo amplification. The sequence of the nucleic acid binding compounds bound to the solid phase will be selected such that they altogether contain a sequence covering the sequence to be determined in the nucleic acid. Within this sequence, the sequences of each nucleic acid binding compound will overlap by one or more bases. In the determination step of this method, it will be detected to which of these partial sequences the nucleic acid will bind, which will only occur (ideally), if a sequence complementary to the partial sequence is contained on a nucleic acid to be determined. Preferably, the evaluation of the result of

the assay will be made by a computer. The computer will thus determine from the partial sequences that are apparently contained in the sequence to be determined, in which series they must have been arranged in the whole nucleic acid. Especially in this kind of assays, the present invention is very helpful, as this approach needs a high
5 number of nucleic acid binding compounds to be fixed on the surface, which can not always take into account the problems occurring with high G-C content. In the present invention, it is therefore possible to sequence even nucleic acids having sequences of low and of high G-C content simultaneously.

- 10 It is apparent to a man skilled in the art that any compound as disclosed herein can to some extent be present as tautomers and salts. These tautomers and salts, preferably the alkali salts, most preferred the sodium salts, are within the definition of the formulae and subject to the present invention.

The present invention is explained in more detail by the following examples:

Examples

5 General

Monomers: Flash chromatography (FC): at 0.5 bar with silica gel 60 (Merck, Darmstadt, Germany). Solvent systems for FC and TLC: CH₂Cl₂-MeOH 85:15 (A), EtOAc-MeOH 3:1 (B), CH₂Cl₂-MeOH 80:20 (C), CH₂Cl₂-HOAc-MeOH 17:1:3 (D), CH₂Cl₂-MeOH 10
9:1 (E), CH₂Cl₂-acetone 85:15 (F). Samples were collected with an UltroRac II fractions collector (LKB Instruments, Sweden). Melting points: Büchi SMP-20 apparatus (Büchi, Switzerland). UV spectra: U-3200 spectrophotometer (Hitachi, Japan). NMR spectra: AC-250 and AMX-500 spectrometers (Bruker, Germany); δ values are relative to internal Me₄Si or external H₃PO₄. Fluorescence spectra were recorded in H₂O on a
15 F-4500 fluorescence spectrophotometer (Hitachi, Japan). Microanalyses were performed by Mikroanalytisches Laboratorium Beller (Göttingen, Germany).

Oligonucleotides: Oligonucleotides were synthesized with a ABI 392 DNA synthesizer (Applied Biosystems, Germany) according to the standard protocol using the "trityl-off" mode, except for the unmodified oligodeoxynucleotides which were synthesized using
20 the "trityl-on" mode. The coupling yields of modified phosphoramidites were generally 95% on average (trityl conductivity monitoring). The detritylated modified oligomers were purified by ion-exchange chromatography on a Dionex Nucleopac PA-100 HPLC column (4 x 250 mm, P/N 043010, Dionex GmbH, Idstein, Germany) using the
25 following gradient: 5 min 5% 0.01 M NaOH/1.5 M aq. LiCl (X) in 0.01 M NaOH (Y); 25 min 5 - 30% Y in X; 10 min 30 - 5% Y in X; 5 min 5% Y in X. Ion-exchange HPLC apparatus: L-4250 UV/VIS detector, L-6250 Intelligent pump and D-2500 integrator (Merck-Hitachi, Germany). The tritylated unmodified oligonucleotides were purified by RP-18 HPLC using the following apparatus and procedure: 250 x 4 mm RP-18
30 column (Merck, Germany); Merck-Hitachi HPLC apparatus consisting of a 655 A-12 liquid chromatograph with a 655 A variable wavelength UV monitor and a D-2000 Chromato-Integrator (Merck-Hitachi, Darmstadt, Germany); gradients of 0.1 M (Et₃NH)OAc (pH 7.0)/MeCN 95:5 (U) and MeCN (V); gradient I: 0 - 50 min 0 - 50% V in U, flow rate 1 mL/min; gradient II: 0 - 20 min 0 - 20% V in U; 20 - 40 min 20 - 40% V
35 in U, flow rate 1 mL/min. Detritylation was performed by treating the purified

oligomers with a 2.5% dichloroacetic acid solution in CH_2Cl_2 (1 mL) for 5 min. After neutralization with Et_3N , evaporation to dryness, followed by co-evaporation with MeOH, the oligomers were again purified by RP-18 HPLC using the above-mentioned device. Gradient: 0 - 30 min 0 - 20 % V in U, 30 - 35 min 20% V in U, 35 - 40 min 20 - 0% V in U, 40 - 45 min 0% V in U. Subsequent desalting for all oligonucleotides was performed on an RP-19 HPLC column (4 x 100 mm) using the apparatus as described above. Solvent for adsorption: H_2O , solvent for desorption: MeOH- H_2O 3:2. General flow rate: 1 mL/min. MALDI-TOF Mass spectra of the oligonucleotides were measured on a home-built apparatus using UV laser irradiation at 337 nm for 3 nsec.

10

The enzymatic hydrolysis of the oligomers was performed as described in *Helv. Chim. Acta* 1998, 81, 1139-1155, but using a flow rate of 0.6 mL/min. Quantification of the constituents was made on the basis of the peak areas, which were divided by the extinction coefficients of the nucleoside (ϵ_{260} values: dA 15400, dC 7300, dG 11400, dT 8800, $z^2\text{A}_d$ 8200). Snake venom phosphodiesterase EC 3.1.15.1, *Crotallus durissus*) and alkaline phosphatase (EC 3.1.3.1, *E. coli*) used for the enzymatic hydrolysis of oligonucleotides were from Roche Diagnostics GmbH.

15

Determination of melting curves and thermodynamics: Absorbance vs. temperature profiles were measured on Cary 1 or 1E spectrophotometers (Varian, Australia) with a Cary thermoelectrical controller. The T_m values were measured in the reference cell with a Pt-100 resistor, and the thermodynamic data (ΔH° , ΔS° , ΔG°_{298}) were calculated with the program MeltWin 3.0. Circular dichroism (CD) spectra were recorded on a Jasco 600 (Jasco, Japan) spectropolarimeter, a thermostatically controlled bath (Lauda RCS-6) in a 1-cm cuvette.

25

Example 1

3-(2-Deoxy- β -D-erythro-pentofuranosyl)-3H-imidazo[2,1-i]purine (1,N⁶-Etheno-2'-deoxyadenosine, 5). 2'-Deoxyadenosine monohydrate (1) (5.0 g, 20 mmol) was dissolved in 1M aq. sodium acetate buffer (pH 4.5 - 5.0, 110 mL) by warming to 40 - 50°C. To the solution chloroacetaldehyde (50% aq. soln, 7.7 mol/L, 25 mL) was added, and the reaction mixture was stirred for 70 h at room temperature. The yellow solution was evaporated to dryness, and the residue was dissolved in MeOH and filtered to remove inorganic salt. After washing with MeOH the combined filtrate and washings

35

were concentrated in vacuo at 40 - 50°C. The residue was applied to FC (silica gel 60H, column: 20 x 6 cm). Elution with CH₂Cl₂-MeOH (85:15) gave a main fraction from which upon evaporation of the solvent and subsequent crystallization from MeOH-EtOAc compd. 5 (3.86 g, 70%) was isolated as colorless crystals. M.p. 138-141°C. TLC (silica gel, EtOAc-MeOH, 3:1): R_f 0.4. UV (MeOH): λ_{max} 275 (7300), 265 (7600), 258 (6600), 229 nm (35700). ¹H-NMR ([D₆]DMSO) δ 2.39 (m, 1H, H_a-C(2')); 2.70 (m, 1H, H_g-C(2')); 3.57 (m, 1H, H_a-C(5')); 3.67 (m, 1H, H_b-C(5')); 3.88 (m, 1H, H-C(4')); 4.43 (m, 1H, H-C(3')); 4.99 (t, 1H, ³J(H,H) = 5.2 Hz, 5'-OH); 5.38 (d, 1H, ³J(H,H) = 3.8 Hz, 3'-OH); 6.47 (pt, 1H, ³J(H,H) = 6.2 Hz, H-C(1')); 7.55 (s, 1H, H-C(11)); 8.07 (s, 1H, H-C(10)); 8.53 (s, 1H, H-C(2)); 9.29 (s, 1H, H-C(8)).

Example 2

1-(2-Deoxy-β-D-erythro-pentofuranosyl)-5-amino-4-(imidazol-2''-yl)-imidazole (6).
Compound 5 (3.85 g, 14 mmol) was treated with 1N aq. NaOH (60 mL) at room temperature overnight. The reaction mixture was adjusted to pH 7 by addition of 2N aq. HCl and concentrated to a syrup. This was dissolved in absolute MeOH, and the precipitated NaCl was filtered off and washed with MeOH. Filtrate and washings were combined and evaporated. The residue was applied to FC (silica gel 60H, column: 20 x 6 cm). Elution with CH₂Cl₂-MeOH (C) afforded a main zone from which compound 6 (2.70 g, 73%) was obtained as a colorless foam which was used for the next reactions without further purification. An analytical sample was crystallized from MeOH-EtOAc to give colorless spherical crystals; m.p. 91-93°C (decomp.). TLC (silica gel, CH₂Cl₂-HOAc-MeOH, 17:1:3): R_f 0.22. UV (MeOH): λ_{max} 271 nm (12800). ¹H-NMR ([D₆]DMSO) δ 2.21 (m, 1H, H_a-C(2')); 2.47 (m, 1H, H_g-C(2')); 3.57 (m, 2H, H₂-C(5')); 3.84 (m, 1H, H-C(4')); 4.36 (m, 1H, H-C(3')); 6.00 (pt, 1H, ³J(H,H) = 6.5 Hz, H-C(1')); 6.60 (br. s, NH₂); 7.13 (s, 2H, H-C(4) + H-C(5)); 7.55 (s, 1H, H-C(2)); 8.16 (s, NH).

Example 3

3-(2-Deoxy- β -D-erythro-pentofuranosyl)-1*H*-diimidazo[1,2-*c*:4',5'-*e*][1,2,3]-triazine (1, *N*⁶-etheno-2-aza-2'-deoxyadenosine, 7). A solution of compound 6 (4.50 g, 17 mmol) in 80% aq. HOAc was treated with sodium nitrite (1.17 g, 17 mmol) in an ice-water bath for 1 h. The reaction mixture was evaporated to a syrup. This was dissolved in H₂O and evaporated repeatedly to remove HOAc. The residue was applied to FC (silica gel 60H, column, 20 x 6 cm). Elution with CH₂Cl₂-MeOH (85:15) afforded compound 7 (2.50 g, 53%) upon evaporation. M.p. 151-152°C (decomp.). TLC (silica gel, CH₂Cl₂-MeOH, 4:1): R_f 0.5. UV (MeOH): λ_{max} 282 (3100), 268 (3200), 238 nm (37900). ¹H-NMR ([D₆]DMSO) δ 2.54 (m, 1H, H_a-C(2')); 2.85 (m, 1H, H_g-C(2')); 3.97 (m, 2H, H₂-C(5')); 4.00 (m, 1H, H-C(4')); 4.50 (m, 1H, H-C(3')); 4.96 (t, 1H, ³J(H,H) = 5.4 Hz, 5'-OH); 5.41 (d, 1H, ³J(H,H) = 4.3 Hz, 3'-OH); 6.69 (pt, 1H, ³J(H,H) = 6.3 Hz, H-C(1')); 7.85 (d, 1H, ³J(H,H) = 1.1 Hz, H-C(11)); 8.75 (d, 1H, ³J(H,H) = 1.1 Hz, H-C(10)); 8.95 (s, 1H, H-C(8)). Anal. calcd. for C₁₁H₁₂N₆O₃ (276.25): C 47.83, H 4.38, N 30.42; found: C 47.71, H 4.32, N 30.32.

Example 4

4-Amino-7-(2-deoxy- β -D-erythro-pentofuranosyl)-7*H*-imidazo[4,5-*d*][1,2,3]-triazine (2-aza-2'-deoxyadenosine, 2). Compound 7 (0.56 g, 2 mmol) was dissolved in 1M aq. sodium acetate buffer (pH 4.0 - 4.5, 120 mL) by warming to 40 - 50°C. To this solution N-bromosuccinimide (2.8 g, 16 mmol) was added, and the reaction mixture was stirred at room temperature overnight. The reaction mixture was evaporated and applied to a Dowex 1x8 ion exchange column (3 x 12 cm, OH⁻ form). Elution with H₂O (250 mL) gave compound 2 (0.19 g, 38%) as colorless needles which decompose above 185°C. The reaction product was identical with an authentic sample in all respects ^[21].

Example 5

7-(2-deoxy- β -D-erythro-pentofuranosyl)-7*H*-imidazo[4,5-*d*][1,2,3]-triazin-4-one (2-aza-2'-deoxyinosine, 3). Compound 2 (19 mg, 0.076 mmol) was dissolved in H₂O and adenosine deaminase (2 μ g, from calf intestine, dissolved in glycerole) was added. The reaction mixture was stirred for 18 h at room temperature until 2 had completely disappeared (UV monitoring) and then evaporated to dryness in a SpeedVac

concentrator. UV (H₂O): λ_{\max} 247 (5500), 290 nm (6200). ¹H-NMR (D₂O): 2.49 (m, 1H, H_a-C(2')); 2.75 (m, 1H, H_B-C(2')); 3.41, 3.50 (2m, 2H, H₂-C(5')); 4.03 (m, 1H, H-C(4')); 4.51 (m, 1H, H-C(3')); 6.43 (pt, 1H, ³J(H,H) = 3.2 Hz, H-C(1')); 8.31 (s, 1H, H-C(8)).

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Example 6

4-(Benzoylamino)-7-(2-deoxy- β -D-erythro-pentofuranosyl)-7H-imidazo[4,5-d][1,2,3]-triazine (8). Compound 2 (125 mg, 0.5 mmol) was co-evaporated twice with anhydrous pyridine. The residue was suspended in anhydrous pyridine and treated with trimethylsilyl chloride (0.5 mL, 4 mmol). After few minutes of stirring a clear solution was formed. The reaction mixture was stirred at room temperature for 2 h. Next, benzoyl chloride (0.25 mL, 2 mmol) was added, and stirring was continued for another 2 h. The reaction mixture was cooled in an ice-water bath, and H₂O (1 mL) was added. After 10 min the reaction mixture was treated with aqueous conc. NH₃ (0.8 mL) and left for additional 30 min. The mixture was then evaporated to dryness, treated with H₂O, and extracted with EtOAc (3 X 20 mL). The combined extracts were dried (Na₂SO₄) and applied onto a silica gel column (3 X 15 cm). Elution was performed with CH₂Cl₂ (150 mL), followed by CH₂Cl₂-MeOH (9 : 1). The nucleoside-containing fractions were evaporated to dryness, and compound 8 was crystallised from MeOH-H₂O to give colorless needles (135 mg, 76%). M.p. 208-210° C (decomp > 170°C). TLC (silica gel, CH₂Cl₂-MeOH 9:1): R_f 0.31. UV: (10% MeOH in water): λ_{\max} 233 (15600), 276 nm (16400). ¹H-NMR ([D₆] DMSO): δ 2.97 (2m, 2H, H₂-C(2')); 3.69 (m, 2H, H₂-C(5')); 4.00 (m, 1H, H-C(4')); 4.57 (m, 1H, H-C(3')); 5.07 (t, 1H, ³J(H,H) = 4.8 Hz, 5'-OH); 5.49 (d, 1H, ³J(H,H) = 4.0 Hz, 3'-OH); 6.72 (t, 1H, ³J(H,H) = 6.5 Hz, H-C(1')); 7.61-7.78 (m, 4H, aromatic-H), 8.16 (d, 2H, aromatic-H); 9.09 (s, 1H, H-C(8)); 11.84 (s, 1H, N-H). Anal. calcd. for C₁₆H₁₆N₆O₄ (356.3): C 52.93, H 4.41, N 23.38; found: C 52.68, H 4.39, N 23.07.

25

Example 7

7-(2-Deoxy- β -D-erythro-pentofuranosyl)-4-[[[(dimethylamino)methylidene]-amino]-
7H-imidazo[4,5-d][1,2,3]-triazine (9a). To a stirred suspension of compound 2 (63 mg,
5 0.25 mmol) in MeOH (5 mL) *N,N*-dimethylformamide dimethylacetal (120 mg, 0.5
mmol) was added. Stirring was continued for 2 h at room temperature. The reaction
mixture was evaporated to dryness, and the residue was adsorbed on silica gel. Flash
chromatography on a silica gel column (3 X 10 cm) with CH₂Cl₂ (100 mL) followed by
CH₂Cl₂-MeOH (9:1) afforded colorless needles (MeOH-H₂O, 65 mg, 85%). M.p. 173-
10 175°C. TLC (silica gel, CH₂Cl₂-MeOH, 9:1): R_f 0.28. UV: (10% MeOH in H₂O): λ_{\max}
234 (13250), 319 nm (29500). ¹H-NMR ([D₆] DMSO): δ 2.84 (2m, 2H, H-C(2')); 3.17,
3.25 (2s, 2H, N-CH₃); 3.60 (m, 2H, H₂-C(5')); 3.92 (m, 1H, H-C(4')); 4.47 (m, 1H, H-
C(3')); 5.05 (t, 1H, ³J(H,H) = 4.9 Hz, 5'-OH); 5.39 (d, 1H, ³J(H,H) = 4.0 Hz, 3'-OH);
6.55 (t, 1H, ³J(H,H) = 6.6 Hz, H-C(1')); 8.79 (s, 1H, N=CH); 9.08 (s, 1H, H-C(8)).
15 Anal. calcd. for C₁₂H₁₇N₇O₃ (307.3): C 46.90, H 5.58, N 31.90; found: C 46.55, H 5.68, N
31.66.

Example 8

20 7-(2-Deoxy- β -D-erythro-pentofuranosyl)-4-[[[(di-isobutylamino)methylidene]-
amino]-7H-imidazo[4,5-d][1,2,3]-triazine (9b). As described for 9a but using *N,N*-di-
isobutylformamide dimethylacetal. Colorless crystals (72%). M.p. 138-140°C. TLC
(silica gel, CH₂Cl₂-MeOH, 9:1): R_f 0.40. UV (10% MeOH in water): λ_{\max} 236 (10100),
325 nm (25850). ¹H-NMR ([D₆]DMSO): δ 0.88, 0.94 (2d, 12 H, CH₃); 1.95, 2.20 (2m,
25 2H, CH); 2.80 (2m, 2H, H₂-C(2')); 3.30-3.74 (m, 6H, H₂-C(5') and 2 CH₂); 4.00 (m,
1H, H-C(4')); 4.45 (m, 1H, H-C(3')); 5.03 (t, 1H, ³J(H,H) = 4.9 Hz, 5'-OH); 5.37 (d,
1H, ³J(H,H) = 4.0 Hz, 3'-OH); 6.54 (t, 1H, ³J(H,H) = 6.4 Hz, H-C(1')); 8.78 (s, 1H,
N=CH); 9.10 (s, 1H, H-C(8)). Anal. calcd. for C₁₈H₂₉N₇O₃ • ½ H₂O (400.5): C 53.99, H
7.55, N 24.48; found: C 53.65, H 7.62, N 24.11.

Example 9

7-(2-Deoxy- β -D-erythro-pentofuranosyl)-4-[[di-n-butylamino)methylidene]-amino}-
7H-imidazo[4,5-d][1,2,3]-triazine (9c). As described for 9a but using N,N-di-n-
5 butylformamide dimethylacetal. Colorless needles (75%). M.p. 107-109° C. TLC (silica
gel, CH₂Cl₂-MeOH, 9:1): R_f 0.42. UV: (10% MeOH in water): λ_{max} 235 (10200), 325 nm
(25700). ¹H NMR ([D₆]DMSO): δ 0.93 (t, 6H, CH₃), 1.33, 1.64, 3.70 (3m, 12H, -CH₂-);
2.45, 2.80 (2m, 2H, H₂-C(2')); 3.60 (m, 2H, H₂-C(5')); 3.92 (m, 1H, H-C(4')); 4.48 (m,
1H, H-C(3')); 5.04 (t, 1H, ³J(H,H) = 5.8 Hz, 5'-OH); 5.39 (d, 1H, ³J(H,H) = 4.0 Hz, 3'-
10 OH); 6.55 (pt, 1H, ³J(H,H) = 6.3 Hz, H-C(1')); 8.78 (s, 1H, N=CH); 9.08 (s, 1H, H-
C(8)). Anal. calcd. for C₁₈H₂₉N₇O₃ (391.5): C 55.23, H 7.47, N 25.05; found: C 55.36, H
7.66, N 24.97.

Example 10

15 7-[2-Deoxy- 5-O-(4,4'-dimethoxytriphenylmethyl)- β -D-erythro-pentofuranosyl]-4-
[[di-n-butylamino)methylidene]amino]-7H-imidazo[4,5-d][1,2,3]-triazine (10a).
Compound 9c (390 mg, 1 mmol) was co-evaporated twice with pyridine, and the oily
residue was dissolved in anhydrous pyridine (6 mL). Next, 4,4'-dimethoxytriphenyl-
20 methyl chloride (450 mg, 1.3 mmol) was added, and the reaction mixture was stirred
at room temperature for 2 h. Thereupon, MeOH (0.2 mL) was added, and stirring
was continued for 15 min. The reaction mixture was poured into 15 mL of an aq. 5%
NaHCO₃ solution, and this was extracted twice with CH₂Cl₂ (30 mL, each). The
combined extracts were dried over Na₂SO₄, evaporated, and the residue was adsorbed
25 on silica gel. This was applied onto a silica gel 60H - column (4 X 14 cm) and
chromatographed with a CH₂Cl₂-acetone gradient (0 \rightarrow 25% of acetone, total volume,
600 mL). The nucleoside-containing fractions were pooled and evaporated to obtained
compound 10a as solid foam (560 mg, 81%). TLC: (silica gel, CH₂Cl₂-acetone, 85:15):
R_f 0.15. ¹H-NMR ([D₆] DMSO): 0.93 (t, 6H, CH₃); 1.34, 1.63, 3.75 (3m, 12H, -CH₂-);
30 2.95 (2m, 2H, H-C(2')); 3.51 (m, 2H, H₂-C(5')); 3.63, 3.69 (2s, 6H, OCH₃); 4.01 (m,
1H, H-C(4')); 4.59 (m, 1H, H-C(3')); 5.45 (d, 1H, ³J(H,H) = 4.1 Hz, 3'-OH); 6.57 (pt,
1H, ³J(H,H) = 6.2 Hz, H-C(1')); 6.60 - 7.30 (m, 13H, phenyl-H); 8.71 (s, 1H, N=CH),
9.07. (s, 1H, H-C(8)). Anal. calcd. for C₃₉H₄₇N₇O₅ (693.8): C 67.51; H 6.83; N 14.13;
found: C 67.15, H 6.82, N 14.13.

Example 11

7-[2-Deoxy- 5-O-(4,4'-dimethoxytriphenylmethyl)- β -D-erythro-pentofuranosyl]-4-
5 {[(di-n-butylamino)methylidene]amino}-7H-imidazo[4,5-d][1,2,3]-triazine-3'-[(2-
cyanoethyl)-N,N-diisopropyl phosphoramidite] (10b). To a solution of compound 10a
(300 mg, 0.43 mmol) in anhydrous CH₂Cl₂ (20 mL) N,N-diisopropylethylamine (145
 μ L, 0.88 mmol) and chloro-(2-cyanoethoxy)-N,N-diisopropylaminophosphine (143 μ L,
0.62 mmol) were added under an Ar atmosphere. After stirring for 20 min at room
temperature, a 5% aq. NaHCO₃ solution (15 mL) was added and the mixture was
10 extracted with CH₂Cl₂ (2 x 30 mL). The organic layer was dried (Na₂SO₄), filtered and
evaporated. FC (silica gel, column 5 x 10 cm, CH₂Cl₂/acetone, 85:15) gave a mixture of
diastereoisomers of the title compound (300 mg, 78%). TLC (silica gel, CH₂Cl₂/acetone,
85:15): R_f 0.71, 0.80. ³¹P-NMR (CDCl₃): 149.962, 150.223.

15 Example 12

5'-O-(4,4'-dimethoxytrityl)-6-N-((di-n-butylamino)methylene)-2-aza-2'-
deoxyadenosine 3'-[(2-Cyanoethyl)-N,N-(diisopropyl)]phosphoramidite (12)
To a solution of 5'-O-(4,4'-dimethoxytrityl)-6-N-((di-n-butylamino)methylene)-2-aza-
20 2'-deoxyadenosine (300 mg, 0.43 mmol) in anh. CH₂Cl₂ (20 ml) (i-Pr)₂EtN (145 μ L, 0.88
mmol) and chloro-(2-cyanoethoxy)(diisopropylamino)phosphine (143 μ L, 0.62 mmol)
were added. After stirring for 20 min at r. t., a 5% aq. NaHCO₃ solution (15 ml) was
added and the mixture was extracted with CH₂Cl₂ (2 x 30 ml). The organic layer was
dried (Na₂SO₄), filtered and evaporated. FC (silica gel, column 5 x 10 cm,
25 CH₂Cl₂/acetone, 85:15) gave a mixture of diastereoisomers NR-411 (300 mg, 77%). TLC
(silica gel, CH₂Cl₂/acetone, 85:15): R_f 0.71, 0.80. ³¹P-NMR (CDCl₃): 149.962, 150.223.

Example 13

30 Synthesis of nucleic acid binding compounds using the monomers of Example 12

The synthesis was performed as outlined in the General section.

Example 14

35

Determination of nucleic acids using probes according to Example 13

Table 1. T_m -Values and Thermodynamic Parameters of Duplex Formation of Oligonucleotides Containing 2-Aza-2'-deoxyadenosine.

5	Oligonucleotide	T_m	ΔH	ΔS	ΔG
		[°C]	[kcal/mol]	[cal/K mol]	[kcal/mol]
10	5'-d(TAGGTCAATACT)				
	3'-d(ATCCAGTTATGA)	46	-82	-230	-10.4
	5'-d(TAGGTC6ATACT)				
	3'-d(ATCCAGTTATGA)	42	-85	-245	-9.2
15	5'-d(TAGGTC6ATACT)				
	3'-d(ATCCAGGTATGA)	46	-83	-236	-9.9
	5'-d(TAGGTC66TACT)				
	3'-d(ATCCAGTTATGA)	37	-76	-219	-7.7
20	5'-d(TAGGTC66TACT)				
	3'-d(ATCCAGAAATGA)	25	-49	-141	-5.7
	5'-d(TAGGTC66TACT)				
	3'-d(ATCCAGCCATGA)	20	-41	-113	-5.5
	5'-d(TAGGTC66TACT)				
	3'-d(ATCCAGGGATGA)	46	-74	-206	-10.1
30	5'-d(TAGGTCAATACT)				
	3'-d(ATCCAGGGATGA)	36	-47	-127	-7.4
	5'-d(TAGGTCGGTACT)				
	3'-d(ATCCAGCCATGA)	54			
35	10 mM Na-cacodylate, 100 mM NaCl, 10 mM MgCl ₂ , pH 7; 5 μ M single strand concentration; 6: $z^2 A_d$.				

The two oligonucleotides 5'-d(TAGGTCAATACT) and 5'-d(AGTATTGACCTA) were constructed to form a stable hybrid with a T_m value of 47°C. This duplex is used as a standard to study the influence of modified bases on the duplex structure and stability. As can be seen from Table 1 the replacement of one central dA - dT by a z^2A_d - dT base pair reduces the T_m of the duplex by 5°; the exchange of two base pairs reveals a decrease of the T_m by 10°. The reduction of duplex stability is obviously independent from the position of base pair replacement: the duplex having two consecutive z^2A_d - dT pairs, exhibits the same T_m as the duplex in which the modified base pairs are separated by three regular ones. Duplex stability is linearly decreased further when the number of z^2A_d - dT base pairs is increased; the oligonucleotide containing four modified pairs exhibits a T_m of only 28°C. This result is in striking contrast to findings on oligonucleotides in which dA residues are replaced by 8-aza-2'-deoxyadenosine; here, the introduction of even four z^8A_d residues instead of dA does not exert any influence on the duplex stability.

As can be seen from table 1, the T_m of an oligonucleotide having at position 7 a G-C base pair (last line in table 1) has a T_m of 54 °C. Replacement of two C's in these base pairs by 2-azaadenine yields a T_m of 46 °C (7 duplex). The same T_m is present when these artificial base pairs are replaced by the natural base pair A-T (first duplex). Moreover, the duplex is having mixed 2-azaadenine/G and A-T base pair (third duplex). From table 1 it can be learned that replacement of one G-C base pair by an artificial base pair of the present invention reduces the T_m by between 3 and 5 °C, preferably 4 °C.

The above described results were found for duplexes with antiparallel chain orientation. The same result, however, was also found for oligonucleotide duplexes with a parallel strand polarity. The chain orientation of naturally-occurring DNA is antiparallel (aps). This orientation can be turned to parallel when the duplex contains isoG_d - dC and or isoMe⁵C_d - dG base pairs (Helv. Chim. Acta. 1997, 80, 73-85). As the pairing of dA with dT is ambiguous any natural DNA can be hybridized in the parallel mode when the second strand contains the bases isoguanine, isocytosine, adenine and thymine. As an example, one duplex is given in Table 1. When in this duplex two dA - dT base pairs are replaced by z^2A_d - dT, a reduction of the T_m value by 10° is determined which is identical with the results for corresponding antiparallel oligonucleotide duplexes.

The T_m data listed in Table 1 display another interesting feature of the base pairing properties of z^2A_d (2). Stimulated by the finding that replacement of a destabilizing central z^2A_d - dT base pair by z^2A_d - dG enhances the T_m value of the oligomer back to the value of the unmodified duplex (T_m 46°C), we investigated the duplex stabilities of
5 oligomers containing mismatches.

For this purpose oligodeoxynucleotides were synthesized in which two central z^2A_d residues are placed opposite to either two dT, dA, dC or dG residues. In all cases, except for z^2A_d - dG - containing duplexes, the T_m value is significantly decreased - most
10 pronounced for the oligomer with two z^2A_d - dC pairs. This oligonucleotide, however, exhibits almost the same value as the unmodified duplex. This prompted us to propose a z^2A_d - dG base pair as shown in FIG 1 and FIG 6 (motif I). Thus, the present invention can also be used in assays where nucleic acids should be discriminated using mismatches.

15

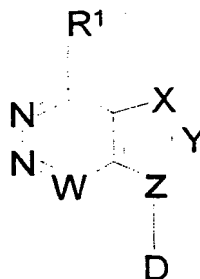
The findings on the peculiar base pairing of 2-aza-2'-deoxyadenosine imply that this nucleoside exhibits similar pairing properties as 2'-deoxyisoguanosine ($isoG_d$), the more so as both show a similar hydrogen bonding donor-acceptor pattern when assuming a keto/H-N(3) tautomeric form of 2'-deoxyisoguanosine (FIG 6, motif II). Indeed, the
20 latter forms a purine-purine base pair with 2'-deoxyguanosine in oligodeoxynucleotides with an antiparallel strand polarity but significantly weaker base pairs with dC and dT, and particularly with dA.

With the aid of these results we anticipate that in parallel oriented oligonucleotides
25 2-aza-2'-deoxyadenosine will form a base pair with $isoG_d$ (FIG 7, motif III) under neutral conditions as well as with protonated dC (FIG 7, motif V) in antiparallel arranged duplexes. On the other hand, with protonated 2'-deoxyisocytidine an antiparallel base pair should be formed following the structural motif VI depicted in FIG 7.

Claims:

1. A nucleic acid binding compound comprising a backbone, said backbone having
 5 attached heterocyclic groups capable of base pairing to natural nucleobases at least
 one of said heterocyclic groups being one of the naturally occurring nucleobases
 characterized in that at least one other of said heterocyclic groups is a group of the
 general formula I

Formula I



10 wherein

W is selected independently from X, Y and Z from the group consisting of N
 and CR²,

15 Z is selected from the group consisting of N and C with the proviso that

- if Z is N, then

20 X independently from W and Y is selected from the group
 consisting of N and CR³, and

Y independently from W and X is selected from the group
 consisting of N and CR⁴,

and the bond between X and Y is a double bond and the bond
 between Y and Z is a single bond, and

25

- if Z is C, then

X is NR^{33} , and

Y is selected from the group consisting of N and CR^4 and

5 the bond between Z and Y is a double bond and the bond between X and Y is a single bond,

R^1 , R^2 , R^3 and R^4 are independently selected from the group consisting of -H, -
halogen, $-\text{OR}^{13}$, $-\text{SR}^{19}$, $-(\text{C}_1-\text{C}_{10})$ -alkyl, $-(\text{C}_2-\text{C}_{10})$ -alkenyl, $-(\text{C}_2-\text{C}_{10})$ -alkynyl,
10 $-\text{NO}_2$, $-\text{NR}^5\text{R}^6$, -cyano, and $-\text{C}(=\text{O})\text{R}^{11}$,

R^{11} is selected from the group consisting of -OH, $-(\text{C}_1-\text{C}_6)$ -alkoxy, $-(\text{C}_6-\text{C}_{22})$ -
aryloxy, and NHR^{12} ,

R^5 , R^6 , R^{12} , R^{13} , R^{19} and R^{33} are selected independently from the group consisting
of -H, $-(\text{C}_1-\text{C}_{10})$ -alkyl, $-(\text{C}_2-\text{C}_{10})$ -alkenyl, $-(\text{C}_2-\text{C}_{10})$ -alkynyl, $-(\text{C}_6-\text{C}_{22})$ -aryl, a
15 protecting group and a reporter group,

r and s are independently of each other an integer of 1 to 18,

D is the position of attachment of the group to the rest of the nucleic acid
binding compound, and

said alkyl, alkenyl and alkynyl being unsubstituted or substituted by one or more
20 moieties selected from the group consisting of -halogen, -SH, $-\text{S}-(\text{C}_1-\text{C}_6)$ -alkyl, $-(\text{C}_1-\text{C}_6)$ -alkoxy, -OH, $-\text{NR}^5\text{R}^6$, $-\text{COR}^{11}$, $-\text{NH}-\text{CONR}^5\text{R}^6$, $-\text{NH}-\text{CSNR}^5\text{R}^6$ and
 $-\text{[O}-(\text{CH}_2)_r\text{]}_s-\text{NR}^5\text{R}^6$.

2. The nucleic acid binding compound of claim 1, wherein the backbone comprises
25 sugar and phosphate moieties.

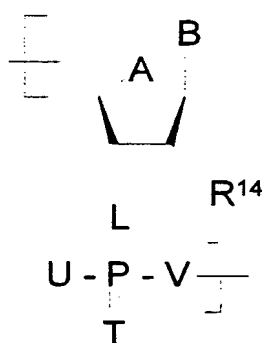
3. A nucleic acid binding compound of claim 2, wherein the sugar configuration is
selected from the group consisting of the α -D-, β -D-, α -L- and β -L-
configurations.

30

4. The nucleic acid binding compound of claim 3, wherein the sugar moiety is a
2'-deoxy- β -D-erythropentofuranosyl moiety.

5. The nucleic acid binding compound of claim 1, wherein R¹ is selected from the group consisting of -SH, -(C₁-C₆)-alkoxy, -(C₂-C₆)-alkylmercapto, -NR⁵R⁶, F and NO₂.
6. The nucleic acid compound of claim 5, wherein R¹ is -NR⁵R⁶.
7. The nucleic acid binding compound of any of claims 1 - 6, wherein the backbone comprises one or more moieties of the general formula II

Formula II



wherein

- 10

A is selected from the group consisting of O, S and N-(C₁-C₁₀)-alkyl,

L is selected from the group consisting of oxy, sulfanediyl and $-NR^{22}-$,

T is selected from the group consisting of oxo, thioxo and selenoxo,

U is selected from the group consisting of -OH, -O-reporter group, -SH, -S

15

reporter group -SeH, -(C₁-C₁₀)-alkoxy, (C₁-C₁₀)-alkyl, -(C₆-C₂₂)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -NR²³R²⁴, and -O-(C₁-C₁₀)-alkyl-O-(C₁-C₁₀)-alkyl-R²⁵, or wherein -NR²³R²⁴ can together with N be a 5-6-membered heterocyclic ring,

V is selected from the group consisting of oxy, sulfanediyl or $-NR^{22}-$,

20

R¹⁴ is selected from the group consisting of -H, -OH, -(C₁-C₁₀)-alkoxy, -(C₂-C₁₀)-alkenyloxy, -halogen, -azido, -O-allyl, -O-alkynyl, and -NH₂,

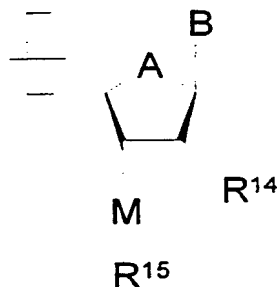
R^{22} is independently selected from the group of -H and -(C₁-C₁₀)-alkyl,

R²³ and R²⁴ are independently selected from the group consisting of -(C₁-C₁₀)-alkyl, -(C₁-C₂₀)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -(C₁-C₆)-alkyl-[NH(CH₂)_c]_d-NR²⁶R²⁷ and a reporter group,

25

- R^{25} is selected from the group consisting of -H, -OH, -halogen, -amino, -(C₁-C₁₈)-alkylamino, -COOH, -CONH₂ and -COO(C₁-C₄)-alkyl and a reporter group,
- R^{26} and R^{27} are independently selected from the group consisting from -H, -(C₁-C₆)-alkyl, and -(C₁-C₄)-alkoxy-(C₁-C₆)-alkyl and a reporter group,
- c is an integer from 2 to 6,
- d is an integer from 0 to 6, and
- B is a moiety of formula I,
- wherein any alkyl, alkenyl and alkynyl can be substituted or unsubstituted.
8. The nucleic acid binding compound of claim 1, wherein R^1 is -NH₂.
 9. The nucleic acid compound of claim 1 containing at least one reporter group.
 10. The nucleic acid binding compound of claim 1, wherein the backbone comprises a moiety of the general formula III

Formula III



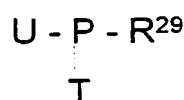
wherein

- A is selected from the group consisting of O, S and N-(C₁-C₆)-alkyl,
- M is selected from the group consisting of oxy, sulfanediyl, -NR²²-, -(C₁-C₁₀)-alkyl-, or -O-(C₁-C₁₀)-alkyl-O-, and -S-(C₁-C₁₀)-alkyl-O- and -NR²²-(C₁-C₆)-alkyl-O-,
- R^{22} is selected from the group of -H, -(C₁-C₁₀)-alkyl, a protecting group and a reporter group,

R^{14} is selected from the group consisting of -H, -OH, -(C₁-C₁₀)-alkoxy, -(C₂-C₁₀)-alkenyloxy, -(C₂-C₁₀)-alkynyloxy, -halogen, -azido, SH, -(C₁-C₁₀)-alkylmercapto and -NH₂,

5 R^{15} is selected from the group consisting of -H, -(C₁-C₆)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₂-C₁₀)-alkynyl, -(C₂-C₁₀)-alkyl-carbonyl, -(C₃-C₁₉)-alkenyl-carbonyl, -(C₃-C₁₉)-alkynyl-carbonyl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, a solid phase and a group of formula IV

Formula IV



wherein

10

T is selected from the group consisting of oxo, thioxo and selenoxo, and

U is selected from the group consisting of -OH, -O-reporter group, -SH, -SeH, -(C₁-C₁₀)-alkoxy, -(C₁-C₁₀)-alkyl, -(C₆-C₂₂)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -NR²³R²⁴, and -O-(C₁-C₁₀)-alkyl-O-(C₁-C₁₀)-alkyl-R²⁵, or wherein NR²³R²⁴ can together with N be a 5-6-membered heterocyclic ring,

15

R²³ and R²⁴ are independently selected from the group consisting of -(C₁-C₁₀)-alkyl, -(C₁-C₂₀)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -(C₁-C₆)-alkyl-[NH(CH₂)_c]_d-NR²⁶R²⁷,

R²⁵ is selected from the group consisting of -H, -OH, -halogen, -amino, -(C₁-C₁₈)-alkylamino, -COOH, -CONH₂ and -COO(C₁-C₄)-alkyl,

20

R²⁶ and R²⁷ are independently selected from the group consisting from -H, -(C₁-C₆)-alkyl, and -(C₁-C₄)-alkoxy-(C₁-C₆)-alkyl

R²⁹ is selected from the group consisting of -OR³⁰ and -SR³⁰,

R³⁰ is selected from the group consisting of -H, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₆-C₂₂)-aryl, a protecting group, a solid phase and a reporter group

25

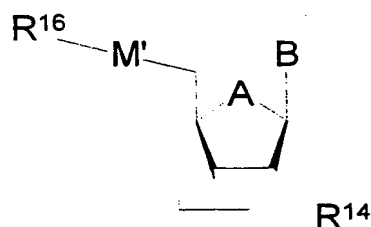
B is the link to a moiety of formula I,

wherein any alkyl, alkenyl and alkynyl can be substituted or unsubstituted.

30

11. The nucleic acid binding compound of any of claims 1, 5 and 6, wherein said backbone comprises a moiety of the formula V

Formula V



5 wherein

- A is selected from the group consisting of O, S and N-(C₁-C₆)-alkyl,
M' is selected from the group consisting of oxy, sulfanediyl, -NR²²-, -(C₁-C₁₀)-
alkyl, or -O-(C₁-C₁₀)-alkyl-O-, and -S-(C₁-C₁₀)-alkyl-O- and -NR²²-(C₁-
10 C₆)-alkyl-O-,
R²² is selected from the group of -H, a protecting group, a reporter group and
-(C₁-C₁₀)-alkyl,
R¹⁴ is selected from the group consisting of -H, -OH, -(C₁-C₁₀)-alkoxy,
-(C₂-C₁₀)-alkenyloxy, -(C₂-C₁₀)-alkynyloxy, -halogen, azido, -SH, -S-(C₁-
15 C₆)-alkylmercapto and NH₂,
R¹⁶ is selected from the group consisting of -H, -(C₁-C₈)-alkyl, -(C₂-C₁₈)-
alkenyl, -(C₂-C₁₈)-alkynyl, -(C₂-C₁₈)-alkyl-carbonyl, -(C₃-C₁₉)-alkenyl-
carbonyl, -(C₃-C₁₉)-alkynyl-alkonyl, -(C₆-C₁₄)-aryl-(C₁-C₈)-alkyl, a
protective group or a compound of formula IV

20

Formula IV

U - P - R²⁹

T

wherein

- T is selected from the group consisting of oxo, thioxo and selenoxo,
U is selected from the group consisting of -OH, -SH, -SeH, -(C₁-C₁₀)-alkoxy,
25 -(C₁-C₁₀)-alkyl, -(C₆-C₂₂)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -NR²³R²⁴, and

-O-(C₁-C₁₀)-alkyl-O-(C₁-C₁₀)-alkyl-R²⁵, wherein NR²³R²⁴ can together with N be a 5-6-membered heterocyclic ring,

R²³ and R²⁴ are independently selected from the group consisting of -(C₁-C₁₀)-alkyl, -(C₁-C₂₀)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -(C₁-C₆)-alkyl-[NH(CH₂)_c]_d-NR²⁶R²⁷,

R²⁵ is selected from the group consisting of -H, -OH, -halogen, -amino, -(C₁-C₁₈)-alkylamino, -COOH, -CONH₂ and -COO(C₁-C₄)-alkyl,

R²⁶ and R²⁷ are independently selected from the group consisting from -H, -(C₁-C₆)-alkyl, and -(C₁-C₄)-alkoxy-(C₁-C₆)-alkyl

R²⁹ is selected from the group consisting of -OR³⁰ and -SR³⁰,

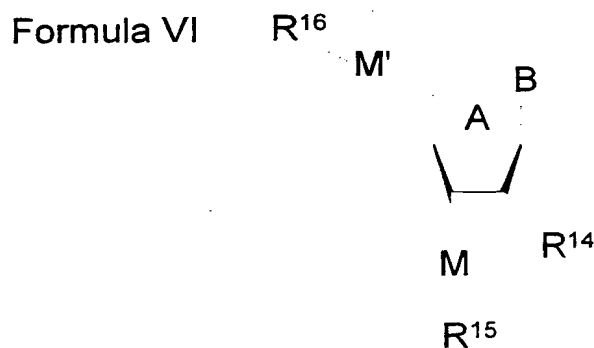
R³⁰ is selected from the group consisting of -H, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₆-C₂₂)-aryl, a protecting group, a solid phase and a reporter group, and

B is the link to a moiety of formula I,

wherein any alkyl, alkenyl and alkynyl can be substituted or unsubstituted.

12. The compound of claim 11, wherein M' is O, R¹⁶ is H and R¹⁴ is selected from the group consisting of -H and -OH.

13. A compound of formula VI

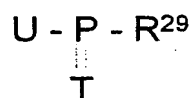


wherein

A is selected from the group consisting of O, S and N-(C₁-C₆)-alkyl,

R¹⁴ is selected from the group consisting of -H, -OH, -(C₁-C₁₀)-alkoxy, O-protecting group, S-protecting group, NH-protecting group, -(C₂-C₁₀)-alkenyloxy, -halogen, -azido, -SH, -(C₁-C₆)-alkylmercapto and -NH₂,
 R¹⁵ and R¹⁶ are independently selected from the group consisting of -H, -(C₁-C₈)-alkyl, -(C₂-C₁₈)-alkenyl, -(C₂-C₁₈)-alkynyl, -(C₂-C₁₈)-alkyl-carbonyl, -(C₃-C₁₉)-alkenyl-carbonyl, -(C₃-C₁₉)-alkynyl-carbonyl, -(C₆-C₁₄)-aryl-(C₁-C₈)-alkyl, a protecting group or a compound of formula IV

Formula IV



wherein

T is selected from the group consisting of oxo, thioxo and selenoxo,
 U is selected from the group consisting of -OH, -SH, -SeH, -(C₁-C₁₀)-alkoxy, -(C₁-C₁₀)-alkyl, -(C₆-C₂₂)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -NR²³R²⁴, and -O-(C₁-C₁₀)-alkyl-O-(C₁-C₁₀)-alkyl-R²⁵, or wherein NR²³R²⁴ can together with N be a 5-6-membered heterocyclic ring,
 R²³ and R²⁴ are independently selected from the group consisting of -(C₁-C₁₀)-alkyl, -(C₁-C₂₀)-aryl, -(C₆-C₁₄)-aryl-(C₁-C₁₀)-alkyl, -(C₁-C₆)-alkyl-[NH(CH₂)_C]_d-NR²⁶R²⁷,
 R²⁵ is selected from the group consisting of -H, -OH, -halogen, amino, -(C₁-C₁₈)-alkylamino, -COOH, -CONH₂ and COO(C₁-C₄)-alkyl,
 R²⁶ and R²⁷ are independently selected from the group consisting from -H, -(C₁-C₆)-alkyl, and -(C₁-C₄)-alkoxy-(C₁-C₆)-alkyl,
 R²⁹ is selected from the group consisting of -OR³⁰ and -SR³⁰,
 R³⁰ is selected from the group consisting of -H, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₆-C₂₂)-aryl, a protecting group, a diphosphate and a reporter group, and
 M and M' are independently selected from the group consisting of oxy, sulfanediyl, -NR²², -(C₁-C₁₀)-alkyl, or -O-(C₁-C₁₀)-alkyl-O-, and -S-(C₁-C₁₀)-alkyl-O- and -NR²²-(C₁-C₆)-alkyl-O-,
 R²² is selected from the group of -H and -(C₁-C₁₀)-alkyl, and

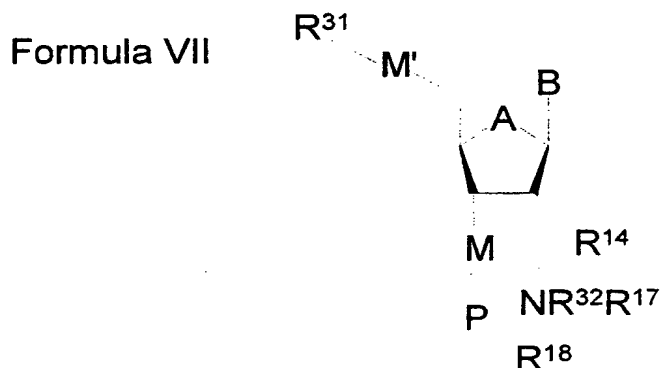
B is a moiety of formula I,

wherein any alkyl, alkenyl and alkynyl can be substituted or unsubstituted, and wherein at least one of R^{15} and R^{16} is not a group of formula IV with the proviso that

MR^{16} , MR^{15} and R^{14} are not each -OH if R^1 is -NH₂ and if either

- W and X and Y and Z is N, or
- W and X and Z is N and Y is CR⁴, or
- W and Y and Z is N and X is CR³.

14. A compound of formula VII



wherein

A is selected from the group consisting of O, S and N-(C₁-C₆)-alkyl, M and M' are independently selected from the group consisting of oxy, sulfanediyl, -NR²², -(C₁-C₁₀)-alkyl, or -O-(C₁-C₁₀)-alkyl-O-, and -S-(C₁-C₁₀)-alkyl-O- and -NR²²-(C₁-C₆)-alkyl-O-,

R²² is selected from the group of -H and -(C₁-C₁₀)-alkyl,

R¹⁴ is selected from the group consisting of -H, -OR³¹, -(C₁-C₁₀)-alkoxy, -(C₂-C₁₀)-alkenyloxy, -(C₂-C₁₀)-alkynyloxy, -halogen, -azido NHR³¹, SR³¹ and -NH₂,

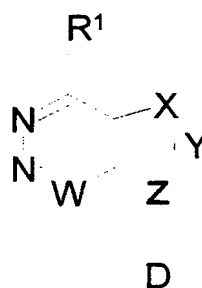
R³¹ is a protecting group or a reporter group,

R³² and R¹⁷ are independently selected from the group consisting of -H, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl and -(C₆-C₂₂)-aryl,

R^{18} is selected from the group consisting of substituted or unsubstituted $-(C_1-C_6)$ -alkyl, unsubstituted $-(C_1-C_6)$ -alkoxy or $-(C_1-C_6)$ -alkoxy substituted one or more times by a group selected from the group consisting of -halogen, p-nitroaryloxy and -cyano, and

5 B is a group of formula I

Formula I



wherein

10

W is selected independently from X, Y and Z from the group consisting of N and CR^2 ,

Z is selected from the group consisting of N and C with the proviso that

15

- if Z is N, then

X independently from W and Y is selected from the group consisting of N and CR^3 , and

Y independently from W and X is selected from the group consisting of N and CR^4 ,

20

and the bond between X and Y is a double bond and the bond between Y and Z is a single bond, and

- if Z is C, then

25

X is NR^{33} , and

Y is selected from the group consisting of N and CR^4 and

the bond between Z and Y is a double bond and the bond between X and Y is a single bond,

- R^1 , R^2 , R^3 and R^4 are independently selected from the group consisting of -H, -halogen, $-OR^{13}$, $-SR^{19}$, $-(C_1-C_{10})$ -alkyl, $-(C_2-C_{10})$ -alkenyl, $-(C_2-C_{10})$ -alkynyl, $-NO_2$, $-NR^5R^6$, -cyano, and $-C(=O)R^{11}$,
- 5 R^{11} is selected from the group consisting of -OH, $-(C_1-C_6)$ -alkoxy, $-(C_6-C_{22})$ -aryloxy, and NHR^{12} ,
- R^5 , R^6 , R^{12} , R^{13} , R^{19} and R^{33} are selected independently from the group consisting of -H, $-(C_1-C_{10})$ -alkyl, $-(C_2-C_{10})$ -alkenyl, $-(C_2-C_{10})$ -alkynyl, $-(C_6-C_{22})$ -aryl, a protecting group and a reporter group,
- 10 r and s are independently of each other an integer of 1 to 18,
- D is the position of attachment of the group to the rest of the nucleic acid binding compound, and
- alkyl, alkenyl and alkynyl being unsubstituted or substituted by one or more moieties selected from the group consisting of -halogen, $-S-(C_1-C_6)$ -alkyl, $-(C_1-C_6)$ -alkoxy, $-NR^5R^6$, $-CO-R^{11}$, $-NH-CO-NR^5R^6$, $-NH-CSNR^5R^6$ and $-[O-(CH_2)_r]_s-NR^5R^6$,
- 15 $-NR^5R^6$,
with the proviso that at least one of R^5 and R^6 of $-NR^5R^6$ is a protecting group.
15. A compound of claim 14, wherein R^1 is not OR^{31} .
- 20 16. A compound of claim 14, wherein said group of formula I contains at least one reporter group.
17. A compound of any of claims 14 and 15, wherein said group of formula I is
25 selected from the group consisting of groups of formula I, wherein either
- W is N, Z is N, Y is N and X is CR^3 , or
 - W is N, Z is C, Y is N and X is CR^3 , or
 - W is N, Z is N, Y is N and X is N.
- 30 18. A compound of claim 5 with the proviso that R^{31} is not H.
19. The binding product of at least one nucleic acid binding compound of any of claims 1 to 11 and a nucleic acid, the nucleic acid binding compound and the

nucleic acid being bound to each other by base pairing in parallel or antiparallel orientation.

20. A method for the determination of a nucleic acid comprising the steps

5

- providing a sample suspected to contain said nucleic acid,
- providing a nucleic acid binding compound of claim 1, which is essentially complementary to a part or all of said nucleic acid,
- contacting said sample with said nucleic acid binding compound under
- 10 conditions for binding said nucleic acid binding compound to said nucleic acid,
- determining the binding product formed from said nucleic acid and said nucleic acid binding compound as a measure of the presence of said nucleic acid.

15

21. A method of claim 20, wherein any groups of formula I in said nucleic acid binding compound of claim 1 is located in said compound as to base pair to a G moiety in said nucleic acid.

20

22. Use of a 2-azapurine in a nucleic acid binding compound as a substitute for cytosine.

23. The use of a 2-azapurine in hybridization reactions of probes with nucleic acids as a base at a position of the probe base pairing with G in the nucleic acid.

25

24. A method for the chemical synthesis of a compound of any of claims 1 to 13 using activated subunits, wherein said subunit contains at least one group of formula I.

25. A method of claim 24, wherein at least one subunit is a compound of any of

30

claims 13 to 21.

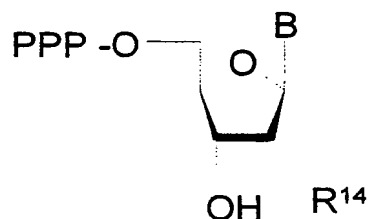
26. A method for the enzymatic synthesis of a compound of any of claims 1 to 13, comprising reacting a triphosphate subunit with a primer using a nucleic acid as a template for the elongation of the primer, wherein the triphosphate subunit

35

contains a heterocyclic group of formula I.

27. A method according to claim 26, wherein said triphosphate subunit has the formula VIII

Formula VIII



5

wherein

PPP is a triphosphate group,

R^{14} is selected from the group consisting of -H, -OH, $-(C_1-C_{10})$ -alkoxy,

10 $-(C_2-C_{10})$ -alkenyloxy, $-(C_2-C_{10})$ -alkynyloxy halogen, -azido and NH_2 , and

B is a group of formula I,

with the proviso that R^{14} is not OH if B is 2-azaadenine.

- 15 28. A method for the determination of the presence or absence of nucleic acids each comprising a particular sequence in a sample comprising the steps

- contacting said sample with a solid phase having immobilized on its surface nucleic acid binding compounds each containing a sequence complementary to one of said particular sequences of said nucleic acids,

20

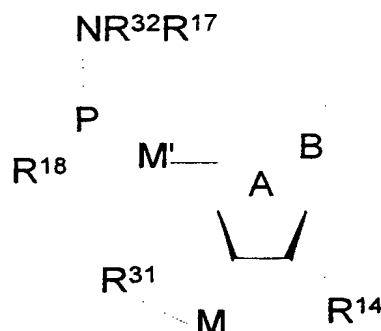
- determining on said solid phase the formation of hybrids containing a nucleic acid with a particular sequence and the nucleic acid binding compound containing the complementary sequence

25 characterized in that said at least one of said nucleic acid binding compounds is a compound according to any of claims 1 to 13.

29. A method for the determination of the presence, absence or amount of a nucleic acid in a sample comprising the steps:

32. A compound of claim 31, wherein R^{14} is -OH.
33. A compound of claim 30, wherein R^{14} is -H and R^1 is not OH.
- 5 34. A compound of general formula IX

Formula IX



wherein

10

A is selected from the group consisting of O, S and N-(C₁-C₆)-alkyl,
M and M' are independently selected from the group consisting of oxy,
sulfanediyl, -NR²², -(C₁-C₁₀)-alkyl, or -O-(C₁-C₁₀)-alkyl-O-, and -S-
(C₁-C₁₀)-alkyl-O- and -NR²²-(C₁-C₆)-alkyl-O-,

15

R²² is selected from the group of -H and -(C₁-C₁₀)-alkyl,

R¹⁴ is selected from the group consisting of -H, -OR³¹, -(C₁-C₁₀)-alkoxy,
-(C₂-C₁₀)-alkenyloxy, -(C₂-C₁₀)-alkynyloxy, -halogen, -azido NHR³¹, SR³¹
and -NH₂,

R³¹ is a protecting group or a reporter group,

20

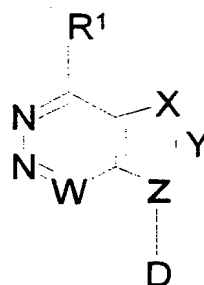
R³² and R¹⁷ are independently selected from the group consisting of -H, -(C₁-C₁₀)-
alkyl, -(C₂-C₁₀)-alkenyl and -(C₆-C₂₂)-aryl,

R¹⁸ is selected from the group consisting of substituted or unsubstituted -(C₁-
C₆)-alkyl, unsubstituted -(C₁-C₆)-alkoxy or -(C₁-C₆)-alkoxy substituted one
or more times by a group selected from the group consisting of -halogen,
p-nitroaryloxy and -cyano, and

25

B is a group of formula I

Formula I



wherein

5 W is selected independently from X, Y and Z from the group consisting of N and CR²,

Z is selected from the group consisting of N and C with the proviso that

- if Z is N, then

10 X independently from W and Y is selected from the group consisting of N and CR³, and

Y independently from W and X is selected from the group consisting of N and CR⁴,

15 and the bond between X and Y is a double bond and the bond between Y and Z is a single bond, and

- if Z is C, then

20 X is NR³³, and

Y is selected from the group consisting of N and CR⁴ and the bond between Z and Y is a double bond and the bond between X and Y is a single bond,

25 R¹, R², R³ and R⁴ are independently selected from the group consisting of -H, -halogen, -OR¹³, -SR¹⁹, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₂-C₁₀)-alkynyl, -NO₂, -NR⁵R⁶, -cyano, and -C(=O)R¹¹,

R¹¹ is selected from the group consisting of -OH, -(C₁-C₆)-alkoxy, -(C₆-C₂₂)-aryloxy, and NHR¹²,

R^5 , R^6 , R^{12} , R^{13} , R^{19} and R^{33} are selected independently from the group consisting of -H, $-(C_1-C_{10})$ -alkyl, $-(C_2-C_{10})$ -alkenyl, $-(C_2-C_{10})$ -alkynyl, $-(C_6-C_{22})$ -aryl, a protecting group and a reporter group,

r and s are independently of each other an integer of 1 to 18,

5 D is the position of attachment of the group to the rest of the nucleic acid binding compound, and

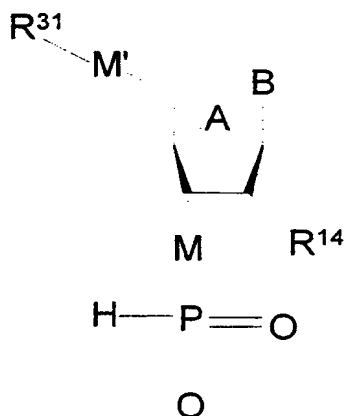
alkyl, alkenyl and alkynyl being unsubstituted or substituted by one or more moieties selected from the group consisting of -halogen, $-S-(C_1-C_6)$ -alkyl, $-(C_1-C_6)$ -alkoxy, $-NR^5R^6$, $-CO-R^{11}$, $-NH-CO-NR^5R^6$, $-NH-CSNR^5R^6$ and $-[O-(CH_2)_r]_s-NR^5R^6$,

10

with the proviso that at least one of R^5 and R^6 of $-NR^5R^6$ is a protecting group.

35. A compound of general formula X

Formula X



15

wherein

M and M' are independently selected from the group consisting of oxy, sulfanediyl, $-NR^{22}$, $-(C_1-C_{10})$ -alkyl, or $-O-(C_1-C_{10})$ -alkyl-O-, and $-S-(C_1-C_{10})$ -alkyl-O- and $-NR^{22}-(C_1-C_6)$ -alkyl-O-,

20

R^{22} is selected from the group of -H and $-(C_1-C_{10})$ -alkyl,

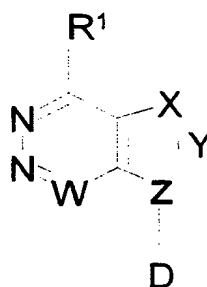
R^{14} is selected from the group consisting of -H, $-OR^{31}$, $-(C_1-C_{10})$ -alkoxy, $-(C_2-C_{10})$ -alkenyloxy, $-(C_2-C_{10})$ -alkynyloxy, -halogen, -azido NHR^{31} , SR^{31} and $-NH_2$,

R^{31} is a protecting group or a reporter group,

25

B is a group of formula I

Formula I



wherein

5 W is selected independently from X, Y and Z from the group consisting of N and CR²,

Z is selected from the group consisting of N and C with the proviso that

- if Z is N, then

10

X independently from W and Y is selected from the group consisting of N and CR³, and

Y independently from W and X is selected from the group consisting of N and CR⁴,

15

and the bond between X and Y is a double bond and the bond between Y and Z is a single bond, and

- if Z is C, then

20

X is NR³³, and

Y is selected from the group consisting of N and CR⁴ and

the bond between Z and Y is a double bond and the bond between X and Y is a single bond,

25

R¹, R², R³ and R⁴ are independently selected from the group consisting of -H, -halogen, -OR¹³, -SR¹⁹, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₂-C₁₀)-alkynyl, -NO₂, -NR⁵R⁶, -cyano, and -C(=O)R¹¹,

R^{11} is selected from the group consisting of -OH, -(C₁-C₆)-alkoxy, -(C₆-C₂₂)-aryloxy, and NHR¹²,

R^5 , R^6 , R^{12} , R^{13} , R^{19} and R^{33} are selected independently from the group consisting of -H, -(C₁-C₁₀)-alkyl, -(C₂-C₁₀)-alkenyl, -(C₂-C₁₀)-alkynyl, -(C₆-C₂₂)-aryl, a

r and s are independently of each other an integer of 1 to 18,

D is the position of attachment of the group to the rest of the nucleic acid binding compound, and

alkyl, alkenyl and alkynyl being unsubstituted or substituted by one or more moieties selected from the group consisting of -halogen, -S-(C₁-C₆)-alkyl,

-(C₁-C₆)-alkoxy, -NR⁵R⁶, -CO-R¹¹, -NH-CO-NR⁵R⁶, -NH-CSNR⁵R⁶ and -[O-(CH₂)_r]_s-NR⁵R⁶.

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Figure 1

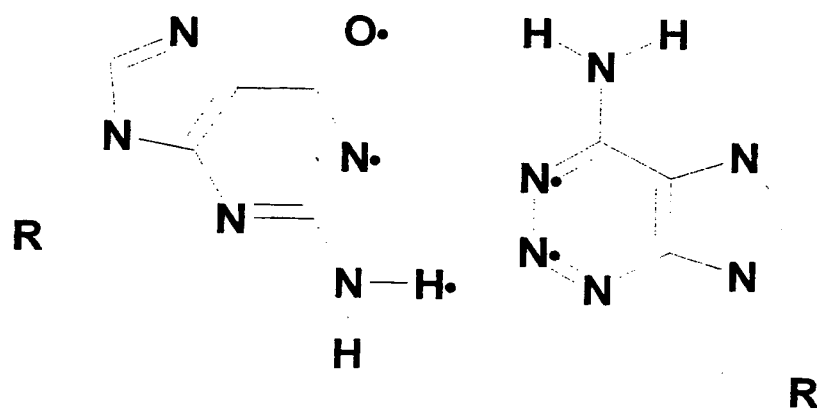
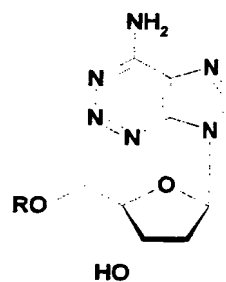
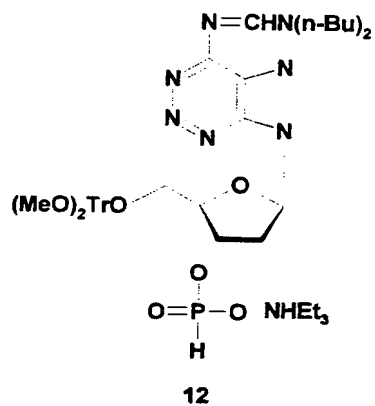


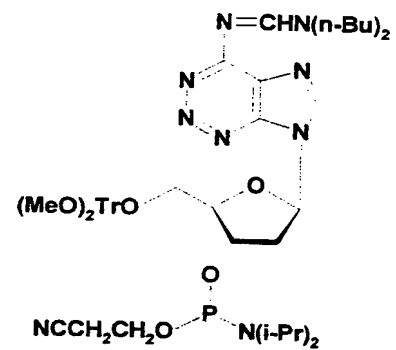
Figure 2



2: R = H
11: R = $P_3O_9H_4$

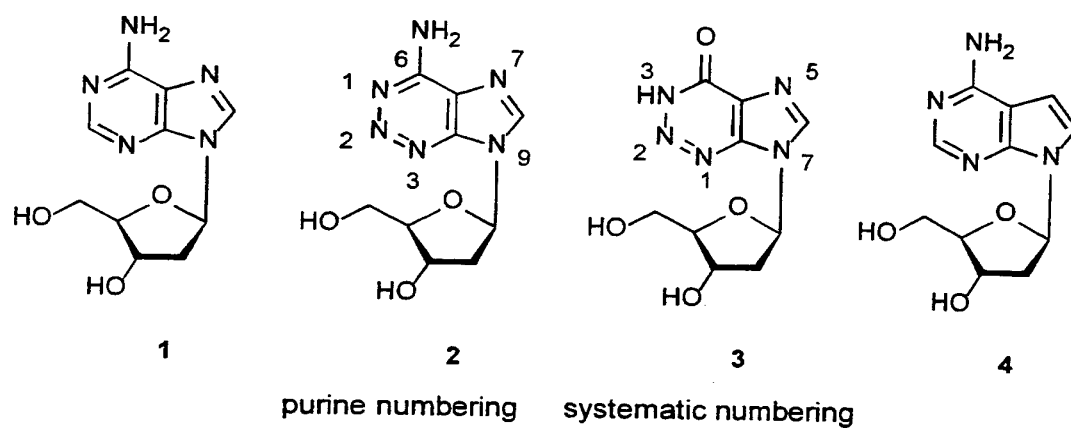


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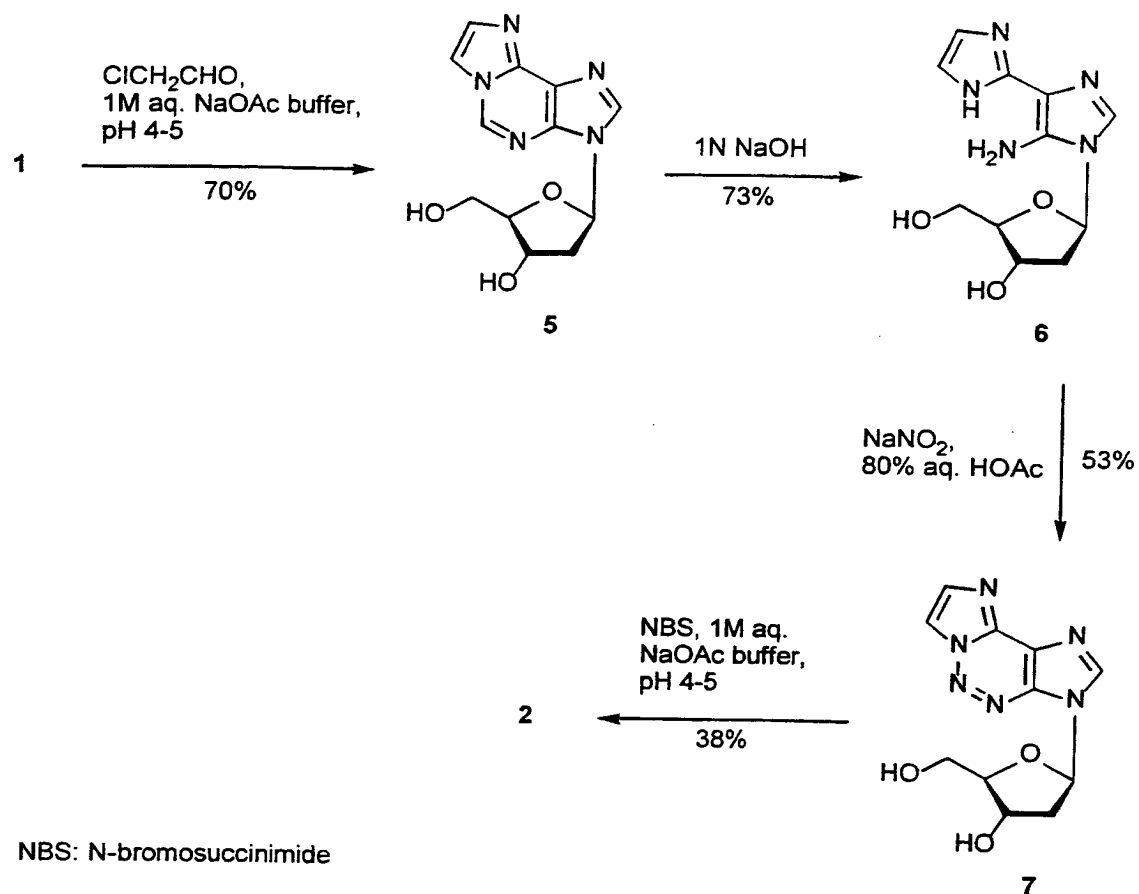
10 b

Figure 3



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Figure 4



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Figure 5

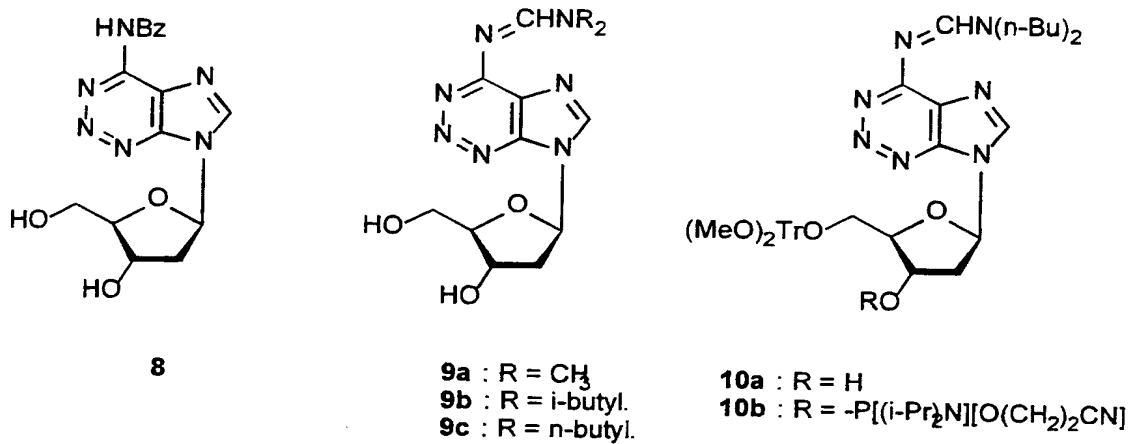
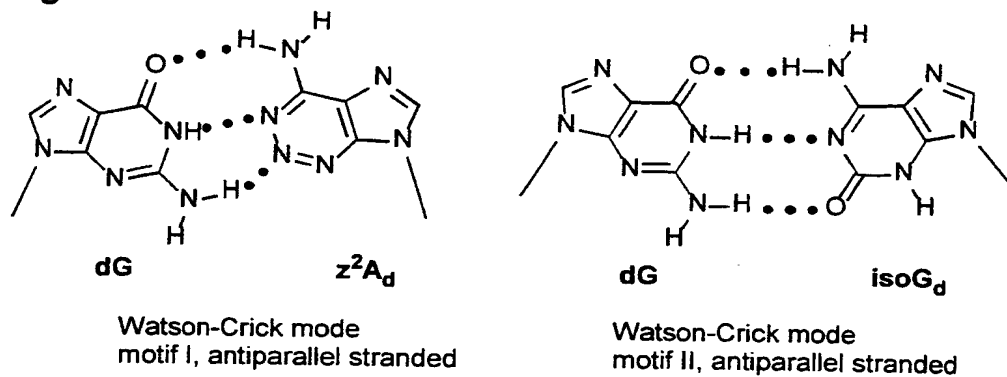
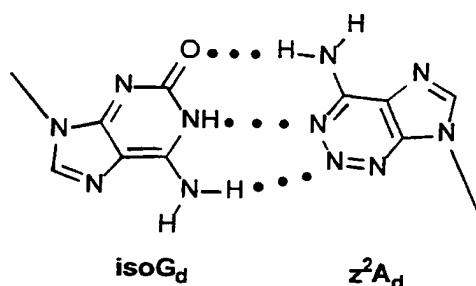


Figure 6

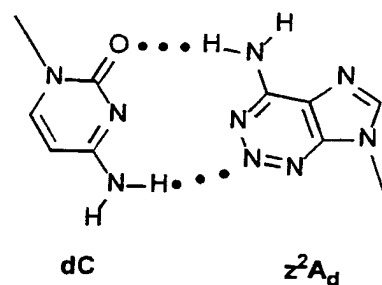


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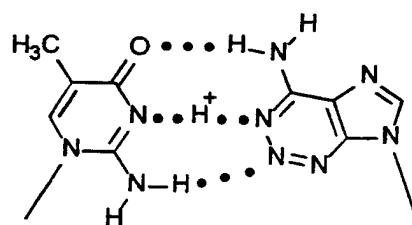
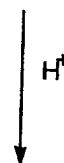
Figure 7



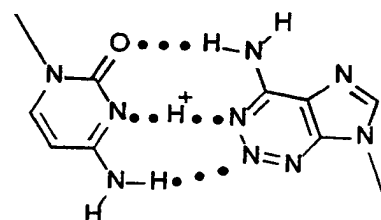
reverse Watson-Crick mode
motif III, parallel stranded



reverse Watson-Crick mode
motif IV, parallel stranded



Watson-Crick mode
motif VI, antiparallel stranded



reverse Watson-Crick mode
motif V, parallel stranded